Supplementary material to:

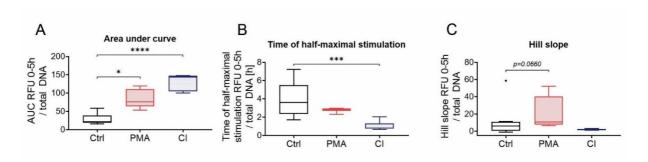
BIO-IMPEDANCE MEASUREMENT ALLOWS DISPLAYING THE EARLY STAGES OF NEUTROPHIL EXTRACELLULAR TRAPS

Caren Linnemann¹, Sascha Venturelli^{2,3}, Franziska Konrad⁴, Andreas K. Nussler¹, Sabrina Ehnert^{1*}

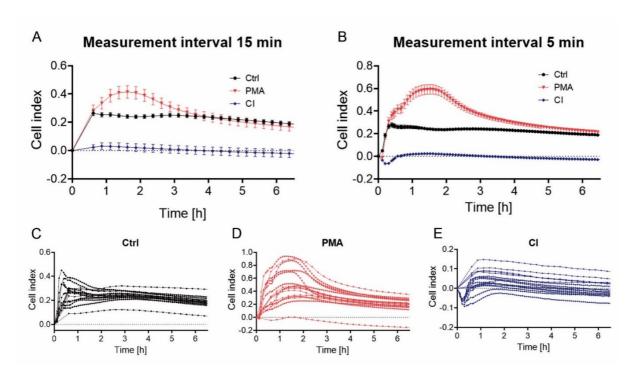
- Siegfried Weller Institute for Trauma Research, BG Unfallklinik Tuebingen, Eberhard Karls Universität Tuebingen, Tuebingen, Germany
- ² Institute of Physiology, Department of Vegetative and Clinical Physiology, University Hospital Tuebingen, Tuebingen, Germany
- Institute of Nutritional Sciences, Department of Nutritional Biochemistry, University of Hohenheim, Stuttgart, Germany
- Department of Anesthesiology and Intensive Care Medicine, University Hospital of Tuebingen, Tuebingen, Germany
- * Corresponding author: Dr. Sabrina Ehnert, Siegfried Weller Institute for Trauma Research at BG Unfallklinik Tuebingen, Eberhard Karls Universität Tuebingen, Schnarrenbergstr. 95, 72076 Tuebingen, Germany; Tel.: +497071 606 1065, Fax.: +49 70 71 606 1978; E-mail: sabrina.ehnert@gmail.com; ORCID: https://orcid.org/0000-0003-4347-1702

http://dx.doi.org/10.17179/excli2020-2868

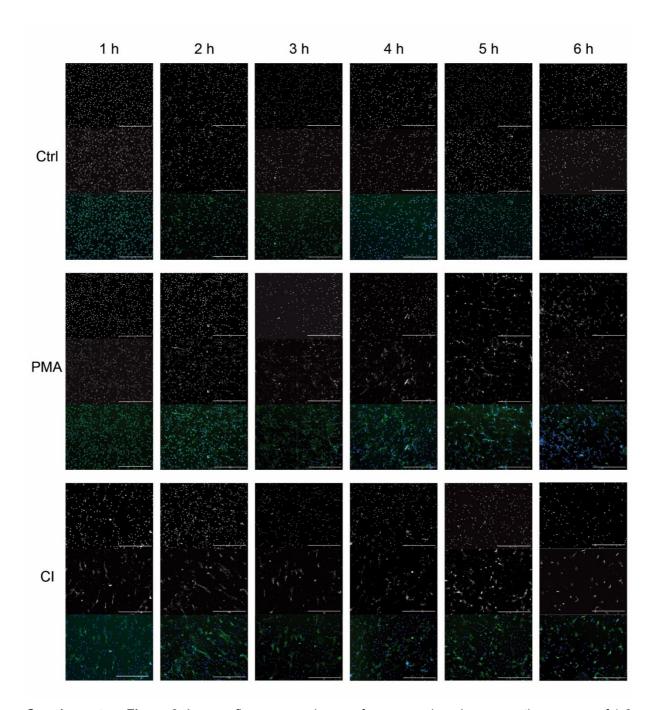
This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/).



Supplementary Figure 1: Sytox Green Assay normalized to total DNA values. Neutrophils were stained with 1 μ M Sytox Green and fluorescence measured over a time of 5 h. Different analysis methods were used to show neutrophil activation by different stimuli. (A) Analysis of DNA release by measurement of area under curve. (B) Analysis of stimulation time by determination of half-maximal stimulation time. (C) Analysis of strength of stimulation by measurement of hill slope. All measured fluorescence data were normalized to total DNA values determined by 1 % Triton-X-100 treated cells. N=15, n=3. Statistical analysis was done by Kruskal-Wallis test followed by Dunn's correction for multiple comparisons. Ctrl: Control, PMA: phorbol 12-myristate 13-acetate, CI: calcium ionophore A23187, AUC: Area under curve, RFU: relative fluorescence unit.



Supplementary Figure 2: Raw data of bio-impedance measurement for the activation of neutrophils. Neutrophils were seeded in eSight E-Plate VIEW 96 and bio-impedance measured for over 6 h. (**A**) Mean values of 8 donors with a measurement interval of 15 min. (**B**) Mean values of 8 donors with a measurement interval of 5 min. (**C**) Values of unstimulated neutrophils of all 15 donors. (**D**) Values of PMA-stimulated neutrophils of all 15 donors. (**E**) Values of CI-stimulated neutrophils of all 15 donors. Mean ± SEM. Ctrl: Control, PMA: phorbol 12-myristate 13-acetate, CI: calcium ionophore A23187.



Supplementary Figure 3: Immunofluorescence images from exemplary donor over time-course of 1-6 h. Example images of one donor over the time course from 1 h to 6 h. Upper panels display nuclear staining with Hoechst 33342 in gray scale, middle panels show myeloperoxidase staining in gray scale. Lower panels display overlay images in color. Scale bar = 400 µm. Blue = Hoechst 33342 (nuclear) staining, green = myeloperoxidase (MPO) staining. Cells were fixed with 4 % formaldehyde at defined time points. Ctrl: Control, PMA: phorbol 12-myristate 13-acetate, CI: calcium ionophore A23187.

Supplementary Table 1: Quality of curve fit analysis of PMA-peak modulation of cell index measurements

Donor	Data points	Adjusted R squared	Standard deviation of residuals
#1	36	0.6767	0.02598
#2		0.6802	0.03737
#3		0.8066	0.01908
#4		0.6311	0.05734
#5		0.8836	0.01447
#6		0.6363	0.03664
#7		0.8313	0.02138
#8		0.3688	0.02199
#9	96	0.9534	0.009359
#10		0.9261	0.02359
#11		0.9482	0.01005
#12		0.9373	0.01229
#13		0.94	0.0282
#14		0.9632	0.01951
#15		0.935	0.03064
#17		0.5936	0.03281
	General formula	$A: Y=B_0 + B_1X + B_2X^2$	1