Original article:

BLADDER CANCER COURSE, FOUR GENETIC HIGH-RISK VARIANTS, AND HISTOPATHOLOGICAL FINDINGS

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ABSTRACT

Urinary bladder cancer, a smoking and occupation related disease, was subject of several genome-wide association studies (GWAS). However, studies on the course of the disease based on GWAS findings differentiating between muscle invasive bladder cancer (MIBC) and non-muscle invasive bladder cancer (NMIBC) are rare. Thus we investigated 4 single nucleotide polymorphisms (SNPs) detected in GWAS, related to the genes coding for TACC3 (transforming, acidic coiled-coil containing protein 3), for FGFR3 (fibroblast growth factor receptor 3), for PSCA (prostate stem cell antigen) and the genes coding for CBX6 (chromobox homolog 6) and APOBEC3A (apolipoprotein B mRNA editing enzyme, catalytic polypeptide-like 3A). This study is based on 712 bladder cancer patients and 875 controls from 3 different case control studies in Germany. The 4 SNPs of interest (PSCA rs2294008 and rs2978974, FGFR3-TACC3 rs798766, and CBX6-APOBEC3A rs1014971) were determined by real-time polymerase chain reaction. The distribution of the 4 SNPs does not vary significantly between cases and controls in the entire study group and in the 3 local subgroups, including two former highly industrialized areas and a region without such history. Also, no significant differences in the bladder cancer subgroups of MIBC and NMIBC were observed. The 4 investigated SNPs do not noticeably contribute differently to the bladder cancer risk for the bladder cancer subgroups of MIBC and NMIBC.

Keywords: Muscle invasive bladder cancer (MIBC), non-muscle invasive bladder cancer (NMIBC), PSCA gene rs2294008 and rs2978974, FGFR3-TACC3 gene region rs798766, CBX6-APOBEC3A gene region rs1014971

INTRODUCTION

Bladder cancer (BC) is the 4th common cancer in males and the 14th common cancer

in females in Germany with an estimated 22.430 men and 7.100 women newly diagnosed in 2014 (Robert Koch Institute, 2018). World-wide, it is the 6th common cancer in

males and the 18th common cancer in females with an estimated 440.864 men and 132.414 women newly diagnosed in 2020 (Sung et al., 2021). This smoking and occupation related disease was a subject of several genome-wide association studies (GWAS; overview in Selinski, 2017; de Maturana et al., 2018). Furthermore, a considerable number of studies investigating the impact of genetically based risk factors on bladder cancer was performed (overview: de Maturana et al., 2018; Golka et al., 2011). Regarding the impact on the course of the disease, multiple loci have been identified, aiming at different endpoints, e.g., T category, grade, recurrence, and mortality yet demonstrating little consensus (Lipunova et al., 2019a).

Thus, we investigated 4 single nucleotide polymorphisms (SNPs) detected in GWAS (Kiemeney et al., 2010; Rothman et al., 2010; Wu et al., 2009; Fu et al., 2012) related to the region of the genes coding for TACC3 and for FGFR3 (transforming, acidic coiled-coil containing protein - fibroblast growth factor receptor 3) located on 4p16.3, the gene coding for PSCA (prostate stem cell antigen) located on 8q24.3, and the region of the genes coding for CBX6 (chromobox homolog 6) and for APOBEC3A (apolipoprotein B mRNA editing enzyme, catalytic polypeptide-like 3A) located on 22q13.1. Furthermore, T category and grading in muscle invasive bladder cancer (MIBC) and non-muscle invasive bladder cancer (NMIBC) and the course of the disease, i.e., the number of the relapses and their histopathological findings, were observed.

The region of the genes coding for TACC3 (transforming, acidic coiled-coil containing protein 3) and for FGFR3 (fibroblast growth factor receptor 3) is located on 4p16.3, a known Huntington disease region (Lee et al., 2012). The reference SNP rs798766(T) is associated with bladder cancer risk (Kiemeney et al., 2010; Meng et al., 2017). rs798766 is located in an intron of TACC3, 70 kb from FGFR3 (Kiemeney et al., 2010). FGFR3 belongs to the family of tyrosine kinases. FGFR3 signaling is involved in development, differentiation, cell survival, migration. angiogenesis. and carcinogenesis (Turner and Grose, 2010). FGFR3 mutations in bladder cancer are mostly activating, followed by gene rearrangements and amplification (Cancer Genome Atlas Research Network, 2014; Helsten et al., 2016). They are mainly detected in genetically stable bladder cancer (van Rhijn et al., 2003) and have been associated with progression in bladder cancer (Parker et al., 2014). FGFR3 gene rearrangements generate constitutively activated and oncogenic FGFR3 kinase protein products, and cellular dependence on these drivers confers sensitivity to selective FGFR inhibition (Williams et al., 2013; Wu et al., 2013; Kim et al., 2018). FGFR alterations are targeted in clinical routine already by the kinase inhibitor erdafitinib, which was approved by FDA in April 2019 (Loriot et al., 2019). Most recently, it was shown in a phase 3 study that erdafitinib is superior to chemotherapy in the therapy of advanced or metastatic bladder cancer in patients with unresectable, advanced or metastatic urothelial cancer and select FGFR3/2 receptor alterations (mutations/fusions) (Loriot et al., 2023) According to the TCGA data base, 15.43 % (29 out of 188) bladder cancer cases provided somatic mutations in their tumors (NIH, 2023).

PSCA (prostate stem cell antigen) was initially identified as a prostate-specific cellsurface antigen (Reiter et al., 1998), but later found to be expressed in many human tissues (Fu et al., 2012). The gene coding for PSCA is located on 8q24.3. This gene encodes a glycosylphosphatidylinositol-anchored cell membrane glycoprotein. It is highly expressed in the prostate and, to lesser extent, in the bladder, placenta, colon, kidney, and stomach. It is up-regulated in many prostate cancers and expressed in cancers of the bladder and pancreas. Its polymorphisms result in an upstream start codon in some individuals (Weizmann Institute of Science, 2022a). rs2978974 is located 10 kb upstream of rs2294008, within an alternative untranslated first exon of PSCA (Fu et al., 2012). The variant rs2294008(T) (Wu et al., 2009; Fu et al.,

2012; Deng et al., 2019; Cui et al., 2019) is associated with bladder cancer risk. In a recent meta-analysis, rs2294008 was associated with bladder cancer (OR = 1.15, 95 % CI 1.11-1.18) (Cui et al., 2019). Furthermore, this SNP was strongly associated with gastric cancer (OR = 1.32, 95 % CI 1.27-1.39). rs2978974 was significant, but to a clearly lower extent, associated with bladder cancer (OR 1.09; 95 % CI 1.03-1.15) (Cui et al., 2019). It is noteworthy, that Fu et al. (2012) reported a significant multiplicative interaction between these two SNPs (P = 0.035). The non-risk allele G of rs2978974 showed strong interaction with nuclear proteins from five cell lines tested, implying a regulatory function. Thus, Fu et al. (2012) concluded that a joint effect of the two PSCA SNPs may be important for bladder cancer susceptibility. According to the TCGA data base, 0.46 % (1 out of 216) bladder cancer cases provided somatic mutations in their tumors (NIH, 2023).

The gene region coding for CBX6 (chromobox homolog 6) and for APOBEC3A (apolipoprotein B mRNA editing enzyme, catalytic polypeptide-like 3A) is located on 22q13.1. This locus is located in a non-genic region approximately 25 kb centromeric of APOBEC3A and 64 kb telomeric of CBX6 (Rothman et al., 2010). APOBEC is a protein family encoded by eleven genes. Subtypes of APOBEC3 can cause specific mutations in RNA and DNA at distinct preferred nucleotide contexts in human cancer (Cao and Wu, 2018). APOBEC enzymes are a major source of mutation in bladder cancer. Tumors enriched for APOBEC mutagenesis have better survival (Glaser et al., 2017). CBX6 is a protein coding gene. It is a component of a Polycomb Group (PcG) multiprotein PRC1 (polycomb repressive complex 1)-like complex, a complex class required to maintain the transcriptionally repressive state of many genes, including Hox genes, throughout development (Vandamme et al., 2011). PcG PRC1 complex acts via chromatin remodeling and modification of histones; it mediates monoubiquitination of histone H2A 'Lys-119', rendering chromatin heritably changed in its expressibility (Weizmann Institute of Science, 2022b). The variant rs1014971(T) is associated with bladder cancer risk (de Maturana et al., 2018; Rothman et al., 2010). CBX gene variants are also associated with risk of other cancers (Li et al., 2020; Lin et al., 2020; Xie et al., 2020). According to the TCGA data base, 0.74 % (3 out of 408) bladder cancer cases provided somatic mutations in APO-BEC3A. Likewise, 0.74 % (3 out of 408) bladder cancer tases provided somatic mutations in CBX6 (NIH, 2023).

MATERIALS AND METHODS

This study is based on 712 bladder cancer patients and 875 controls from 3 different case control studies: Evangelic Hospital, Paul-Gerhardt Foundation Lutherstadt Wittenberg, Lutherstadt Wittenberg, Germany (203 cases and 210 controls (Roth et al., 2012)), St.-Josefs-Hospital Dortmund-Hoerde, Dortmund, Germany (156 cases and 237 controls (Ovsiannikov et al., 2012)), and Rheinland Klinikum Lukaskrankenhaus Neuss, Neuss, Germany (353 cases and 428 controls (Höhne et al., 2017)). Controls were from the same respective hospitals without a known history of cancer. Enrolment of patients was as follows: Lutherstadt Wittenberg from December 1995 to January 1999, Dortmund from July 2009 to December 2010, and Neuss from June 2009 to November 2011. Follow-up derived from medical records was as follows: Lutherstadt Wittenberg from September 2008 to June 2009, Dortmund from May 2012 to August 2012, Neuss from July 2013 to February 2014 (Selinski et al. 2016; Roth et al., 2012).

Analytical methods

DNA of cases and controls was isolated from venous blood samples according to standard methods in the Dortmund Institute (Selinski et al., 2017). The 4 SNPs of interest (PSCA rs2294008 and rs2978974, gene region FGFR3-TACC3 rs798766, and gene region CBX6-APOBEC3A rs1014971) were detected by real-time polymerase chain reaction (rt–PCR) using a TaqMan[®] assay (Saravana Devi et al., 2008; Selinski et al., 2017). The missings for the detected SNPs for cases and controls respectively were as follows: 0 and 0 for rs2294008, 2 and 1 for rs2978974, 0 and 0 for rs798766, and 0 and 4 for rs1014971.

Statistical methods

Hardy-Weinberg Equilibrium was checked using a chi-squared test with one degree of freedom. Chi-squared tests were used testing differences between categorial variables, logistic regression was used to compute Likelihood Ratio and Wald tests as well as odds ratios (OR) and 95 % confidence intervals (95 % CI). The analysis was conducted searching for differences between cases vs controls, recurrent vs non-recurrent and MIBC vs NMIBC and stratified for gender, invasiveness, and smoking habits. False discovery rate (FDR) was used for multiple testing correction.

The studies in the 3 departments of urology were approved by the ethics committee of the Dortmund Institute (24/2009, 31/2009, 50/2011, 74/2014) and by the Institutional Review Board.

RESULTS

The total study group contains 3 subgroups from different regions in Germany with comparable characteristics regarding gender and age among cases as well as among controls. In bladder cancer patients, the portions of the described characteristics were as follows: Total study group: 79 % male, median age 70.5 years. Wittenberg: 86 % male, median age 65.4 years. Dortmund: 75 % male, median age 71.0 years. Neuss: 77 % male, median age 73.2 years. Detailed information, including that of the controls is presented in Table 1 and 2.

A description of the clinical parameters of the study group is presented in Table 3a, b. As expected, in the combined group of muscle invasive bladder cancer (MIBC) and non-muscle invasive bladder cancer (NMIBC), significant differences were observed regarding the smoking habits. A total of 16 % of the male patients, in contrast to 49 % of the female patients were non-smokers (p < 0.0001). The portions of the different T categories did not differ significantly between male and female patients.

		Age								
Study group	Status	Min	Q1*	Median	Q3**	Max	Mean	SD	n	Missing
Total group	case	20.10	62.60	70.50	77.13	93.90	69.30	10.92	712	0
Total group	control	20.50	57.13	69.15	77.10	100.00	65.26	16.23	882	0
Wittenberg	case	20.10	58.90	65.40	71.45	91.30	64.60	11.37	203	0
Wittenberg	control	29.40	61.48	67.80	76.50	91.20	67.78	10.17	212	0
Dortmund	case	43.80	64.08	71.00	78.48	89.20	70.81	9.85	156	0
Dortmund	control	21.70	59.50	70.90	79.10	100.00	68.27	14.66	237	0
Neuss	case	26.10	66.30	73.20	78.30	93.90	71.33	10.30	353	0
Neuss	control	20.50	49.90	69.00	76.60	92.70	62.39	18.79	433	0
NMIBC	case	20.10	62.60	70.00	76.90	93.20	68.95	11.03	581	0
NMIBC	control	20.50	57.13	69.15	77.10	100.00	65.26	16.23	882	0
MIBC	case	41.80	63.60	72.40	78.70	93.90	70.85	10.30	131	0
MIBC	control	20.50	57.13	69.15	77.10	100.00	65.26	16.23	882	0

Table 1: Age distribution in the total study group of bladder cancer cases and the subgroups of nonmuscle invasive bladder cancer (NMIBC) and muscle-invasive bladder cancer (MIBC), and controls

*First quartile **Third quartile

Male patients showed a higher percentage of NMIBC (82 % vs 79 %), whereas female patients showed a higher percentage of MIBC (21 % vs 18 %). Grading differed significantly between the genders in 2 subgroups

(Table 3b). There was no significant genderrelated difference in the distribution of NMIBC and MIBC (Table 3a). A stratification of the clinical parameters for NMIBC and MIBC is presented in Table 3b.

Table 2: Gender distribution in th	e total study group of bladder	cancer cases and the	subgroups of non-
muscle invasive bladder cancer ((NMIBC) and muscle-invasive	bladder cancer (MIB	C), and controls

Gender											
Study group	Status	Total (n)	Male (n)	Male (%)	Female (n)	Female (%)					
Total group	case	712	564	79%	148	21%					
Total group	control	882	655	74%	227	26%					
Wittenberg	case	203	175	86%	28	14%					
Wittenberg	control	212	177	83%	35	17%					
Dortmund	case	156	117	75%	39	25%					
Dortmund	control	237	181	76%	56	24%					
Neuss	case	353	272	77%	81	23%					
Neuss	control	433	297	69%	136	31%					
NMIBC	case	581	464	80%	117	20%					
NMIBC	control	882	655	74%	227	26%					
MIBC	case	131	100	76%	31	24%					
MIBC	control	882	655	74%	227	26%					

Table 3: Description of the total study group of bladder cancer (BC) cases (**a**) and the subgroups of non-muscle invasive bladder cancer (NMIBC) and muscle-invasive bladder cancer (MIBC), stratified for gender, smoking habits, T category, and grading (**b**).

а	Variable	Male (%)	Male (n)	Fe- male (%)	Fe- male (n)	Chi- squared test (P)	LR test (P)	Wald test (P)	OR	95 % CI
BC	non- smokers	16%	91	49%	73	<0.0001	<0.0001	reference		
BC	former smokers	58%	325	28%	41			<0.0001	0.16	0.10-0.24
BC	current smokers	25%	142	22%	33			<0.0001	0.29	0.18-0.47
BC	unknown	1%	6	1%	1			0.1500	0.21	0.01-1.25
BC	total	100%	564	100%	148					
BC	Та	42%	237	45%	67	0.4905	0.5137	reference		
BC	P*	2%	11	2%	3			0.9570	0.96	0.21-3.20
BC	T1	38%	216	32%	47			0.2175	0.77	0.51-1.16
BC	T2	12%	65	16%	23			0.4216	1.25	0.71-2.14
BC	T3-4	6%	35	5%	8			0.6091	0.81	0.34-1.75
BC	total	100%	564	100%	148					
BC	G1	30%	168	38%	56	0.3050	0.3252	reference		
BC	G2	39%	222	35%	52			0.1056	0.70	0.46-1.08
BC	G3	30%	169	26%	39			0.1181	0.69	0.43-1.09
BC	Gx	1%	5	1%	1			0.6443	0.60	0.03-3.82
BC	total	100%	564	100%	148					
BC	NMIBC	82%	464	79%	117	0.4040	0.3748	reference		
BC	MIBC	18%	100	21%	31			0.3695	1.23	0.77-1.91
BC	total	100%	564	100%	148					

*Papillary neoplasm (according to an older WHO classification, applied by the examining pathologist); bold: significant at level $\alpha < 0.05$

b	Variable	Male (n)	Male (%)	Female (n)	Female (%)	Chi- squared	LR test (P)	Wald test (P)	OR	95 % CI
NMIBC	non-	79	17%	56	48%	<0.0001	<0.0001	reference		
	smokers									
NMIBC	former smokers	264	57%	36	31%			<0.0001	0.19	0.12-0.31
NMIBC	current smokers	118	25%	24	21%			<0.0001	0.29	0.16-0.50
NMIBC	unknown	3	1%	1	1%			0.5182	0.47	0.02-3.78
NMIBC	total	464	100%	117	100%					
NMIBC	Та	237	51%	67	57%	0.4242	0.4611	reference		
NMIBC	P*	11	2%	3	3%			0.9570	0.96	0.21-3.20
NMIBC	T1	216	47%	47	40%			0.2175	0.77	0.51-1.16
NMIBC	total	464	100%	117	100%					
NMIBC	G1	168	36%	56	48%	0.0556	0.0821	reference		
NMIBC	G2	210	45%	39	33%			0.0120	0.56	0.35-0.88
NMIBC	G3	84	18%	21	18%			0.3190	0.75	0.42-1.30
NMIBC	Gx	2	0%	1	1%			0.7426	1.50	0.07- 15.95
NMIBC	total	464	100%	117	100%					
MIBC	non smokers	12	12%	17	55%	<0.0001	<0.0001	reference		
MIBC	former smokers	61	61%	5	16%			<0.0001	0.06	0.02-0.18
MIBC	current smokers	24	24%	9	29%			0.0144	0.26	0.09-0.75
MIBC	unknown	3	3%	0	0%			0.9903	0.00	NA 9.17x1070
MIBC	total	100	100%	31	100%					
MIBC	T2	65	65%	23	74%	0.3883	0.3337	reference		
MIBC	T3-4	35	35%	8	26%			0.3430	0.65	0.25-1.55
MIBC	total	100	100%	31	100%					
MIBC	G2	12	12%	13	42%	0.0012	0.0013	reference		
MIBC	G3	85	85%	18	58%			0.0006	0.20	0.08-0.50
MIBC	Gx	3	3%	0	0%			0.9904	0.00	NA 1.10x1071
MIBC	total	100	100%	31	100%					

*Papillary neoplasm (according to an older WHO classification, applied by the examining pathologist); bold: significant at level $\alpha < 0.05$

The frequency of the 4 investigated SNPs in the combined group of NMIBC and MIBC cases and in controls, additionally stratified for the 3 local subgroups, is presented in Table 4.

The distribution of the 4 investigated SNPs, stratified for NMIBC and MIBC cases does not vary significantly between the 3 local subgroups, including two formerly highly industrialized areas with industries associated with elevated bladder cancer risk (Dortmund (Ovsiannikov et al., 2012) and Lutherstadt Wittenberg (Roth et al., 2012)) and a region with no industries with known bladder cancer risk (Neuss (Höhne et al., 2017)). No significant differences between cases and controls and between the subgroups of NMIBC and MIBC bladder cancer were observed (Table 5).

The subgroup of cases with progression stratified for the histopathological findings for T category and grading are presented in Table 6. Due to the small number of cases with progression, a statistical evaluation of the impact of the 4 investigated SNPs was not performed. **Table 4:** Frequency of the double mutated (MM), heterozygous (MN) and double native (NN) status of the 4 investigated single nucleotide polymorphisms (SNPs) in the total group of bladder cancer cases and in controls stratified for the 3 local subgroups

Study	SNP	Gene	Status	Total	MM	MM	MN	MN	NN	NN
group				<u>(n)</u>	<u>(n)</u>	(%)	<u>(n)</u>	(%)	<u>(n)</u>	(%)
Total group	rs2294008	PSCA	case	712	182	26%	358	50%	172	24%
Total group	rs2294008	PSCA	control	875	238	27%	436	50%	201	23%
Wittenberg	rs2294008	PSCA	case	203	48	24%	110	54%	45	22%
Wittenberg	rs2294008	PSCA	control	210	61	29%	110	52%	39	19%
Dortmund	rs2294008	PSCA	case	156	44	28%	71	46%	41	26%
Dortmund	rs2294008	PSCA	control	237	65	27%	115	49%	57	24%
Neuss	rs2294008	PSCA	case	353	90	25%	177	50%	86	24%
Neuss	rs2294008	PSCA	control	428	112	26%	211	49%	105	25%
Total group	rs2978974	PSCA	case	710	87	12%	335	47%	288	41%
Total group	rs2978974	PSCA	control	874	92	11%	399	46%	383	44%
Wittenberg	rs2978974	PSCA	case	203	20	10%	104	51%	79	39%
Wittenberg	rs2978974	PSCA	control	209	22	11%	91	44%	96	46%
Dortmund	rs2978974	PSCA	case	156	17	11%	68	44%	71	46%
Dortmund	rs2978974	PSCA	control	237	28	12%	105	44%	104	44%
Neuss	rs2978974	PSCA	case	351	50	14%	163	46%	138	39%
Neuss	rs2978974	PSCA	control	428	42	10%	203	47%	183	43%
Total group	rs798766	FGFR3- TACC3	case	712	435	61%	250	35%	27	4%
Total group	rs798766	FGFR3- TACC3	control	875	556	64%	291	33%	28	3%
Wittenberg	rs798766	FGFR3- TACC3	case	203	113	56%	79	39%	11	5%
Wittenberg	rs798766	FGFR3- TACC3	control	210	131	62%	71	34%	8	4%
Dortmund	rs798766	FGFR3- TACC3	case	156	97	62%	55	35%	4	3%
Dortmund	rs798766	FGFR3- TACC3	control	237	133	56%	95	40%	9	4%
Neuss	rs798766	FGFR3- TACC3	case	353	225	64%	116	33%	12	3%
Neuss	rs798766	FGFR3- TACC3	control	428	292	68%	125	29%	11	3%
Total group	rs1014971	CBX6- APOBEC3A	case	712	318	45%	321	45%	73	10%
Total group	rs1014971	CBX6- APOBEC3A	control	871	380	44%	382	44%	109	13%
Wittenberg	rs1014971	CBX6- APOBEC3A	case	203	94	46%	81	40%	28	14%
Wittenberg	rs1014971	CBX6- APOBEC3A	control	211	90	43%	95	45%	26	12%
Dortmund	rs1014971	CBX6- APOBEC3A	case	156	69	44%	71	46%	16	10%
Dortmund	rs1014971	CBX6- APOBEC3A	control	236	98	42%	104	44%	34	14%
Neuss	rs1014971	CBX6- APOBEC3A	case	353	155	44%	169	48%	29	8%
Neuss	rs1014971	CBX6- APOBEC3A	control	424	192	45%	183	43%	49	12%

Table 5: Frequency of double mutated (MM), heterozygous (MN) and double native (NN) status of the 4 investigated single nucleotide polymorphisms (SNPs) in bladder cancer (BC) cases, controls and in the BC cases stratified for non-muscle invasive (NMIBC) and muscle invasive (MIBC) bladder cancer

Group	SNP	MM (n)	MN (n)	NN (n)	Chi-squared test P	FDR* corrected P
BC case	rs2294008	182	358	172	0.9198	0.9864
BC control	rs2294008	238	436	201	0.9864	0.9864
NMIBC case	rs2294008	148	284	149	0.6330	0.8440
NMIBC control	rs2294008	238	436	201	0.9864	0.9864
MIBC case	rs2294008	34	74	23	0.1483	0.8173
MIBC control	rs2294008	238	436	201	0.9864	0.9864
BC case	rs2978974	87	335	288	0.5306	0.8296
BC control	rs2978974	92	399	383	0.4619	0.8173
NMIBC case	rs2978974	75	269	236	0.9549	0.9864
NMIBC control	rs2978974	92	399	383	0.4619	0.8173
MIBC case	rs2978974	12	66	52	0.2160	0.8173
MIBC control	rs2978974	92	399	383	0.4619	0.8173
BC case	rs798766	435	250	27	0.2611	0.8173
BC control	rs798766	556	291	28	0.2032	0.8173
NMIBC case	rs798766	353	204	24	0.4768	0.8173
NMIBC control	rs798766	556	291	28	0.2032	0.8173
MIBC case	rs798766	82	46	3	0.3382	0.8173
MIBC control	rs798766	556	291	28	0.2032	0.8173
BC case	rs1014971	318	321	73	0.5877	0.8296
BC control	rs10 1 4971	380	382	109	0.4277	0.8173
NMIBC case	rs1014971	260	260	61	0.7915	0.9864
NMIBC control	rs1014971	380	382	109	0.4277	0.8173
MIBC case	rs1014971	58	61	12	0.5759	0.8296
MIBC control	rs1014971	380	382	109	0.4277	0.8173

*FDR false discovery rate

Relapse*	T category, grading at progression												
	T2	Т3	T4	TxM1	G2	G3	Gx						
R1	13	3	0	2	4	13	1						
R2	3	1	0	0	1	3	0						
R3	1	2	1	0	1	2	1						
R4	1	0	0	0	0	0	1						
Relapse*	Combined T category and grading at progression												
	T2 G2	T2 G3	T2 Gx	T3 G3	T4 Gx	TxM1 G3	TxM1 Gx						
R1	4	9	0	3	0	1	1						
R2	1	2	0	1	0	0	0						
R3	1	0	0	2	1	0	0						
R4	0	0	1	0	0	0	0						
Relapse*			T catego	ory and grad	ling at progr	ession							
		G2	G3	Gx									
R1-R4	T2	6	11	1									
R1-R4	T3	0	6	0									
R1-R4	T4	0	0	1									
R1-R4	TxM1	0	1	1									

Table 6: T category, staging and grading at progression in the investigated bladder cancer cases

*Tumors resected 3 months after prior resection were considered as relapses.

DISCUSSION

Since the first papers on the impact of polymorphism on the metabolism of the antitubercular agent isoniazid (INH) in the 1950s (Hughes et al., 1954; Mitchell et al., 1957), a considerable number of studies have been published on the impact of genetic polymorphisms on bladder cancer risk (overview: Golka et al., 2011). However, with the first paper on genome-wide association studies on bladder cancer risk (Kiemeney et al., 2008), a new era started, showing the impact of genetics on bladder cancer of loci, mostly not associated with bladder cancer before (Golka et al., 2011).

To date, about 40 single nucleotide polymorphisms are shown to be associated with bladder cancer risk, mostly within the range between 1.1 and 1.5, mostly identified and/or confirmed in GWAS (Selinski, 2017; de Maturana et al., 2018; Koutros et al., 2023). This is much lower than the risk conferred by the portion of N-acetyltransferase 2 (NAT2) slow acetylators reported in the first studies on bladder cancer risk due to high exposure to highly carcinogenic aromatic amines in the chemical industry at that time (Lewalter and Miksche, 1992; Cartwright et al., 1982; Golka et al., 2002) or conferred by the portion of GSTM1 negatives reported formerly in highly industrialized areas (Golka et al., 1998; Hung et al., 2004).

The first study on variants detected in GWAS on bladder cancer course was published by Roth et al. (2012), showing an association of longer relapse-free survival in GSTT1 positives (HR = 0.60, 95 % CI = 0.42–0.87) in the formerly highly industrialized area of Lutherstadt Wittenberg. GSTM1 negatives tended to be associated with a better prognosis, whereas the NAT2 risk alleles rs710521(A) and rs9642880(T), detected in the first GWAS on bladder cancer (Kiemeney et al., 2008) tended to be associated with a poor prognosis. Since then, only few studies on the impact of genetic polymorphisms detected in GWAS on the course of the bladder cancer disease have been conducted (Grotenhuis et al., 2014; Lipunova et al.,

2019b), most possibly due to the enormous logistic effort. Suitable candidates for an impact on the course of the disease are those genetic polymorphisms and/or loci, which have a proven influence on bladder cancer risk (Roth et al., 2012). Another promising approach is the investigation of the effect of SNP combinations or, even better, a large panel of SNPs as recently applied for the determination of the risk to contract bladder cancer (Koutros et al., 2023). Although our group has experience with the impact of SNP combinations on bladder cancer risk regarding smoking habits (Selinski et al., 2017), we decided not to apply this approach for this study because the available case numbers are by far too small when two or more polymorphisms are combined.

It is now established for a long time that bladder cancer is no more one tumor entity, but two: Non-muscle invasive bladder cancer (NMIBC) and muscle-invasive bladder cancer (MIBC) (Knowles and Hurst, 2015). It may be noted, that genetic analysis has resulted in a new biological classification for MIBC discriminating 6 different subtypes (Kamoun et al., 2020). Similar activities for NMIBC are undergoing (Marzouka et al., 2022). Due to the very different course of these two tumor entities, we decided to investigate the impact on the subgroups of NMIBC and the clinically much more important MIBC separately in larger study group now including in addition to Roth et al. (2012), patients from two other studies (Ovsiannikov et al., 2012; Höhne et al., 2017). The 4 investigated SNPs showed no noticeable differences in the frequency between the NMIBC and MIBC subgroups. This is in line with the findings of Grotenhuis et al. (2014) for PSCA gene related rs2294008, TACC3 - FGFR gene region related rs798766, and CBX6 - APO-BEC3A gene region related rs1014971. This may be due to the minimal impact of a single SNP. Thus, in future studies an impact of SNP combinations should be investigated on large study groups, as this procedure had successfully shown that a 4 SNP combination confers increased bladder cancer risk particularly in

never smokers (Selinski et al., 2017). Recently, this approach was applied by a GWAS using a 24 SNP marker panel (Koutros et al., 2023). Another promising approach would be to investigate the impact of selected SNPs and/or their combinations in the clinically relevant group of symptomatic patients showing micro and/or gross hematuria.

Author contributions

Conceptualization T.K., S.S., T.O., J.G.H., and K.G., methodology S.S., M.B., and J.R., software S.S., laboratory analysis M.B., J.R., data analysis T.K., S.S., investigation T.K., S.S., T.O., and K.G., resources F.V., O.M., T.O., and J.G.H., data curation E.R., D.O., H.G., D.B., S.H., and S.S., writing-original draft preparation T.K., S.S., M.B., and K.G., writing-review and editing T.K., T.O., and K.G,. visualization T.K., K.G., supervision F.V., O.M., T.O., and J.G.H, project administration J.G.H., K.G. All authors have read and agreed to the published version of the manuscript.

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Institutional review board statement

The study was conducted in accordance with the Declaration of Helsinki and approved by the Institutional Review Board and by the Ethics Committee of Leibniz Research Centre for Working Environment and Human Factors at TU Dortmund (IfADo).

Informed consent statement

Informed consent was obtained from all subjects involved in the study.

Data availability statement

The data are available on reasonable request from the corresponding author.

Conflicts of interest

The authors declare no conflict of interest.

REFERENCES

Cancer Genome Atlas Research Network. Comprehensive molecular characterization of urothelial bladder carcinoma. Nature. 2014;507(7492):315-22. doi: 10.1038/nature12965.

Cao W, Wu W. Apolipoprotein B mRNA editing enzyme, catalytic polypeptide-like gene expression, RNA editing, and microRNAs regulation. Methods Mol Biol. 2018;1699:75-81. doi: 10.1007/978-1-4939-7435-1_5.

Cartwright RA, Glashan RW, Rogers HJ, Ahmad RA, Barham-Hall D, Higgins E, et al. Role of N-acetyl-transferase phenotypes in bladder carcinogenesis: a pharmacogenetic epidemiological approach to bladder cancer. Lancet. 1982;2(8303):842-5. doi: 10.1016/s0140-6736(82)90810-8.

Cui H, Tang M, Zhang M, Liu S, Chen S, Zeng Z, et al. Variants in the PSCA gene associated with risk of cancer and nonneoplastic diseases: systematic research synopsis, meta-analysis and epidemiological evidence. Carcinogenesis. 2019;40:70-83. doi: 10.1093/carcin/bgy151.

de Maturana EL, Rava M, Anumudu C, Sáez O, Alonso D, Malats N. Bladder cancer genetic susceptibility. A systematic review. Bladder Cancer. 2018;4:215-26. doi: 10.3233/BLC-170159.

Deng S, Ren ZJ, Jin T, Yang B, Dong Q. Contribution of prostate stem cell antigen variation rs2294008 to the risk of bladder cancer. Medicine (Baltimore). 2019;98:e15179. doi: 10.1097/MD.000000000015179.

Fu YP, Kohaar I, Rothman N, Earl J, Figueroa JD, Ye Y, et al. Common genetic variants in the PSCA gene influence gene expression and bladder cancer risk. Proc Natl Acad Sci U S A. 2012;109:4974-9. doi: 10.1073/pnas.1202189109.

Glaser AP, Fantini D, Wang Y, Yu Y, Rimar KJ, Podojil JR, et al. APOBEC-mediated mutagenesis in urothelial carcinoma is associated with improved survival, mutations in DNA damage response genes, and immune response. Oncotarget. 2017;9:4537-8. doi: 10.18632/oncotarget.23344.

Golka K, Bandel T, Schlaefke S, Reich SE, Reckwitz T, Urfer W, et al. Urothelial cancer of the bladder in an area of former coal, iron, and steel industries in Germany: a case-control study. Int J Occup Environ Health. 1998;4:79-84. doi: 10.1179/oeh.1998.4.2.79.

Golka K, Prior V, Blaszkewicz M, Bolt HM. The enhanced bladder cancer susceptibility of NAT2 slow acetylators towards aromatic amines: a review considering ethnic differences. Toxicol Lett. 2002;128: 229-41. doi: 10.1016/s0378-4274(01)00544-6.

Golka K, Selinski S, Lehmann ML, Blaszkewicz M, Marchan R, Ickstadt K, et al. Genetic variants in urinary bladder cancer: collective power of the "wimp SNPs". Arch Toxicol. 2011;85:539-54. doi: 10.1007/s00204-011-0676-3.

Grotenhuis AJ, Dudek AM, Verhaegh GW, Witjes JA, Aben KK, van der Marel SL, et al. Prognostic relevance of urinary bladder cancer susceptibility loci. PLoS One 2014;9:e89164. doi: 10.1371/journal.pone.0089164.

Helsten T, Elkin S, Arthur E, Tomson BN, Carter J, Kurzrock R. The FGFR landscape in cancer: analysis of 4.853 tumors by next-generation sequencing. Clin Cancer Res. 2016;22:259-67. doi: 10.1158/1078-0432.CCR-14-3212.

Höhne S, Gerullis H, Blaszkewicz M, Selinski S, Hengstler JG, Otto T, et al. N-acetyltransferase 1*10 genotype in bladder cancer patients. J Toxicol Environ Health 2017;80:417-22. A. doi: 10.1080/10937404.2017.1304727.

Hughes HB, Biehl JP, Jones AP, Schmidt LH. Metabolism of isoniazid in man as related to the occurrence of peripheral neuritis. Am Rev Tuberc. 1954;70:266-73. doi: 10.1164/art.1954.70.2.266.

Hung RJ, Boffetta P, Brennan P, Malaveille C, Hautefeuille A, Donato F, et al. GST, NAT, SULT1A1, CYP1B1 genetic polymorphisms, interactions with environmental exposures and bladder cancer risk in a high-risk population. Int J Cancer. 2004;110:598-604. doi: 10.1002/ijc.20157.

Kamoun A, de Reyniès A, Allory Y, Sjödahl G, Robertson AG. Seiler R. et al. A consensus molecular classification of muscle-invasive bladder cancer. Eur Urol. 2020;77:420-33. doi: 10.1016/j.eururo.2019.09.006.

Kiemeney LA, Thorlacius S, Sulem P, Geller F, Aben KK, Stacey SN, et al. Sequence variant on 8q24 confers susceptibility to urinary bladder cancer. Nat Genet. 2008;40:1307-12. doi: 10.1038/ng.229.

Kiemeney LA, Sulem P, Besenbacher S, Vermeulen SH, Sigurdsson A, Thorleifsson G, et al. A sequence variant at 4p16.3 confers susceptibility to urinary bladder cancer. Nat Genet. 2010;42:415-9. doi: 10.1038/ng.558.

Kim YS, Kim K, Kwon GY, Lee SJ, Park SH. Fibroblast growth factor receptor 3 (FGFR3) aberrations in muscle-invasive urothelial carcinoma. BMC Urol. 2018;18:68. doi: 10.1186/s12894-018-0380-1.

Knowles MA, Hurst CD. Molecular biology of bladder cancer: new insights into pathogenesis and clinical diversity. Nat Rev Cancer. 2015;15:25-41. doi: 10.1038/nrc3817.

Koutros S, Kiemeney LA, Pal Choudhury P, Milne RL, Lopez de Maturana E, Ye Y, et al. Genome-wide association study of bladder cancer reveals new biological and translational insights. Eur Urol. 2023;84:127-37. doi: 10.1016/j.eururo.2023.04.020.

Lee JM, Gillis T, Mysore JS, Ramos EM, Myers RH, Hayden MR, et al. Common SNP-based haplotype analysis of the 4p16.3 Huntington disease gene region. Am J Hum Genet. 2012;90:434-44. doi. 10.1016/j.ajhg.2012.01.005.

Lewalter J, Miksche L. Empfehlungen zur arbeitsmedizinischen Prävention expositions- und dispositionsbedingter Arbeitsstoff-Beanspruchungen. Verh Dt Ges Arbeitsmed. 1992;31:135–9.

Li X, Gou J, Li H, Yang X. Bioinformatic analysis of the expression and prognostic value of chromobox family proteins in human breast cancer. Sci Rep. 2020; 10:17739. doi: 10.1038/s41598-020-74792-5.

Lin K, Zhu J, Hu C, Bu F, Luo C, Zhu X, et al. Comprehensive analysis of the prognosis for chromobox family in gastric cancer. J Gastrointest Oncol. 2020;11: 932-51. doi: 10.21037/jgo-20-208.

Lipunova N, Wesselius A, Cheng KK, van Schooten FJ, Cazier JB, Bryan RT, et al. Systematic review: genetic associations for prognostic factors of urinary bladder Biomark Cancer 2019a;11: cancer. 1179299X19897255. doi: 10.1177/1179299X19897255.

Lipunova N, Wesselius A, Cheng KK, van Schooten FJ, Bryan RT, Cazier JB et al. Genome-wide association study for tumour stage, grade, size, and age at diagnosis of non-muscle-invasive bladder cancer. Eur Urol Oncol. 2019b:2:381-9. doi: 10.1016/j.euo.2018.08.020

Loriot Y, Necchi A, Park SH, Garcia-Donas J, Huddart R, Burgess E, et al. Erdafitinib in locally advanced or metastatic urothelial carcinoma. N Engl J Med. 2019; 381:338-48. doi: 10.1056/NEJMoa1817323.

Loriot Y, Matsubara N, Park SH, Huddart RA, Burgess EF, Houede N, et al. Phase 3 THOR study: Results of erdafitinib (erda) versus chemotherapy (chemo) in patients (pts) with advanced or metastatic urothelial cancer (mUC) with select fibroblast growth factor receptor alterations (FGFRalt). J Clin Oncol. 2023;41(17, Suppl):LBA4619. doi: 10.12020/CO0.2022.41.17

10.1200/JCO.2023.41.17_suppl.LBA4619.

Marzouka NA, Eriksson P, Bernardo C, Hurst CD, Knowles MA, Sjödahl G, et al. The Lund Molecular Taxonomy applied to non-muscle-invasive urothelial carcinoma. J Mol Diagn. 2022;24:992-1008. doi: 10.1016/j.jmoldx.2022.05.006.

Meng XY, Shi MJ, Chen JF, Liao Y, Hu BW, Hireche A. Association between the TACC3 rs798766 polymorphism and risk of urinary bladder cancer: a synthesis based on current evidence. Dis Markers. 2017;2017: 7850708. doi: 10.1155/2017/7850708.

Mitchell R, Bell JC. Clinical implications of isoniazid blood levels in pulmonary tuberculosis. N Engl J Med. 1957;257:1066-70. doi: 10.1056/NEJM195711282572202.

NIH, National Cancer Institute. GDC data portal <u>https://portal.gdc.cancer.gov/.</u> 2023. (accessed 12 June 2023).

Ovsiannikov D, Selinski S, Lehmann ML, Blaszkewicz M, Moormann O, Haenel MW, et al. Polymorphic enzymes, urinary bladder cancer risk, and structural change in the local industry. J Toxicol Environ Health A. 2012;75:557-65. doi: 10.1080/15287394.2012.675308.

Parker BC, Engels M, Annala M, Zhang W. Emergence of FGFR family gene fusions as therapeutic targets in a wide spectrum of solid tumours. J Pathol. 2014;232:4–15. doi: 10.1002/path.4297.

Reiter RE, Gu Z, Watabe T, Thomas G, Szigeti K, Davis E, et al. Prostate stem cell antigen: a cell surface marker overexpressed in prostate cancer. Proc Natl Acad Sci U S A. 1998;95:1735-40. doi: 10.1073/pnas.95.4.1735.

Robert Koch Institute and the Association of Population-based Cancer Registries in Germany. Cancer in Germany 2013/2014. 11th ed (pp 104-7). Berlin: Robert Koch Institute, 2018. doi: 10.25646/5889.

Roth E, Selinski S, Schikowsky C, Seidel T, Volkert F, Blaszkewicz M, et al. Bladder cancer survival in a former industrial area in Saxony-Anhalt, Germany. J Toxicol Environ Health A. 2012;75:1216-25. doi: 10.1080/15287394.2012.709168. Rothman N, Garcia-Closas M, Chatterjee N, Malats N, Wu X, Figueroa JD, et al. A multi-stage genome-wide association study of bladder cancer identifies multiple susceptibility loci. Nat Genet. 2010;42:978-84. doi: 10.1038/ng.687.

Saravana Devi S, Vinayagamoorthy N, Agrawal M, Biswas A, Biswas R, Naoghare P, et al. Distribution of detoxifying genes polymorphism in Maharastrian population of central India. Chemosphere. 2008;70:1835– 9. doi: 10.1016/j.chemosphere.2007.08.008.

Selinski S. Discovering urinary bladder cancer risk variants: Status quo after almost ten years of genome-wide association studies. EXCLI J. 2017;16:1288-96. doi: 10.17179/excli2017-1000.

Selinski S, Bürger H, Blaszkewicz M, Otto T, Volkert F, Moormann O, et al. Occupational risk factors for relapse-free survival in bladder cancer patients. J Toxicol Environ Health A. 2016;79:1136-43. doi: 10.1080/15287394.2016.1219606.

Selinski S, Blaszkewicz M, Ickstadt K, Gerullis H, Otto T, Roth E, et al. Identification and replication of the interplay of four genetic high-risk variants for urinary bladder cancer. Carcinogenesis. 2017;38:1167-79. doi: 10.1093/carcin/bgx102.

Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2021;71:209-49. doi: 10.3322/caac.21660.

Turner N, Grose R. Fibroblast growth factor signalling: from development to cancer. Nat Rev Cancer. 2010;10: 116-29. doi: 10.1038/nrc2780.

van Rhijn BW, Vis AN, van der Kwast TH, Kirkels WJ, Radvanyi F, Ooms EC, et al. Molecular grading of urothelial cell carcinoma with fibroblast growth factor receptor 3 and MIB-1 is superior to pathologic grade for the prediction of clinical outcome. J Clin Oncol. 2003;21:1912-21. doi: 10.1200/JCO.2003.05.073.

Vandamme J, Völkel P, Rosnoblet C, Le Faou P, Angrand PO. Interaction proteomics analysis of polycomb proteins defines distinct PRC1 complexes in mammalian cells. Mol Cell Proteomics. 2011;10:M110.002642. doi: 10.1074/mcp.M110.002642.

Weizmann Institute of Science. GeneCards. The Human Gene Database. PSCA Gene. Prostate Stem Cell Antigen. <u>https://www.genecards.org/cgibin/carddisp.pl?gene=PSCA</u>. 2022a. (accessed 27 December 2022). Weizmann Institute of Science. Gene Cards. The Human Gene Database. CBX6 Gene (Protein Coding) Chromobox 6. <u>https://www.genecards.org/cgibin/carddisp.pl?gene=CBX6</u>. 2022b. (accessed 27 December 2022).

Williams SV, Hurst CD, Knowles MA. Oncogenic FGFR3 gene fusions in bladder cancer. Hum Mol Genet. 2013;22:795–803. doi: 10.1093/hmg/dds486.

Wu X, Ye Y, Kiemeney LA, Sulem P, Rafnar T, Matullo G, et al. Genetic variation in the prostate stem cell antigen gene PSCA confers susceptibility to urinary bladder cancer. Nat Genet. 2009;41:991-5. doi: 10.1038/ng.421. Wu YM, Su F, Kalyana-Sundaram S, Khazanov N, Ateeq B, Cao X, et al. Identification of targetable FGFR gene fusions in diverse cancers. Cancer Discov. 2013;3:636–47. doi: 10.1158/2159-8290.CD-13-0050.

Xie X, Ning Y, Long J, Wang H, Chen X. Diverse CBX family members as potential prognostic biomarkers in non-small-cell lung cancer. FEBS Open Bio. 2020;10:2206-15. doi: 10.1002/2211-5463.12971.