

Original article:

**ENRICHMENT ANALYSIS AND CHROMOSOMAL DISTRIBUTION
OF GOUT SUSCEPTIBLE LOCI IDENTIFIED BY
GENOME-WIDE ASSOCIATION STUDIES**

Mostafa Saadat 

Department of Biology, School of Science, Shiraz University, Shiraz 71467-13565, Iran;
Tel: +98-71-36137432; Fax: +98-71-32280926; E-mail: saadat@shirazu.ac.ir

<https://dx.doi.org/10.17179/excli2023-6481>

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0/>).

ABSTRACT

Gout is an inherited and common inflammatory arthritic disease. Many researchers will identify polymorphic loci of gout susceptibility by conducting genome-wide association studies (GWAS). In the present study, the enrichment analysis and chromosomal distribution were performed using predicted polymorphic loci associated with gout risk. The polymorphic loci associated to gout were obtained from the GWAS database. Overall, this database contains 64,806 gout patients and 2,856,174 controls. Gene ontology functional annotation and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway analysis were performed by using the Enrichr online server. A total of 110 common polymorphic protein-coding loci associated with gout risk were identified and included in the analysis. The results of the KEGG analysis showed that the gout-associated loci were mainly related to ABC transporters, endocrine and other factor-regulated calcium reabsorption, and gastric acid secretion pathways. The gene ontology analysis showed that the biological processes of the gout-associated loci were vascular transport, transport across the blood-brain barrier, positive regulation of transporter activity, and positive regulation of transcription by RNA polymerase II. The top cellular component was the external side of the apical plasma membrane. Statistical analysis revealed that the human chromosome segments 1q22, 4p16.1, 6p21.1-p21.2, 11q13.1-q13.2, 12q13.11-q13.3, and 12q24.1 had significantly bearing higher numbers of gout susceptibility loci.

Keywords: Chromosome, enrichment analysis, gout, gene ontology

INTRODUCTION

Gout is a common inflammatory arthritic disease. It is caused by the deposition of monosodium urate crystals in articular and non-articular structures. Hyperuricemia (elevated blood urate) is the major risk factor for the development of gout. It is often associated with other conditions such as hypertension, cardiovascular disease, diabetes, dyslipidemia, obesity, chronic kidney disease, and kidney stones. Epidemiological studies have reported that gout has an incidence of 0.6-2.9 per 1000 person-years and a prevalence of 0.68-3.90 % in adults (Dalbeth et al., 2021).

Several familial aggregation studies and comparisons of monozygotic and dizygotic

twins for hyperuricemia, renal clearance of urate, and gout have shown that these traits are multifactorial, with significant heritability (Emmerson et al., 1992; Wilk et al., 2000; Bleyer and Hart, 2006; Voruganti et al., 2009; Krishnan et al., 2012; Kuo et al., 2015). This means that both genetic and non-genetic environmental factors are involved in the pathogenesis of these disorders.

Many researchers will identify the genetic elements of gout susceptibility by conducting genome-wide association studies (GWAS) (Sulem et al., 2011; Lai et al., 2012; Shin et al., 2012; Köttgen et al., 2013; Li et al., 2015; Matsuo et al., 2016; Nakayama et al., 2017, 2020; Chen et al., 2018; Jing et al., 2018; Lee

et al., 2019, 2022; Kawamura et al., 2019; Tin et al., 2019; Backman et al., 2021; Dönertaş et al., 2021; Fitzgerald et al., 2022; Jiang et al., 2021; Sandoval-Plata et al., 2021; Toyoda et al., 2022; Lin et al., 2023; Sumpter et al., 2023) or by examining the association between common genetic polymorphisms and gout risk in case-control studies (Dong et al., 2015; Lee et al., 2017; Zou et al., 2018; Kawaguchi et al., 2021).

Today, enrichment analysis (also called gene set enrichment analysis, functional enrichment analysis, or pathway enrichment analysis) is a popular method for analyzing gene/protein sets that is essentially developed using complex statistical analysis methods. These analyses are used to identify classes of genes or proteins that are overrepresented in a large set of genes or proteins. In other words, enrichment analysis is a statistical method for determining enriched or depleted groups of genes or proteins (Subramanian et al., 2005).

In the present study, the enrichment analysis was performed using predicted polymorphic loci associated with gout risk, and the chromosomal distribution of the associated loci was constructed to identify the non-random chromosomal segments associated with gout.

METHODS

Search for gout associated loci

The polymorphic loci associated with gout were retrieved from the Genome Wide Association Studies (GWAS) database (<https://www.ebi.ac.uk/gwas>) on August 10, 2023 using gout as a keyword.

Enrichment analysis

Because enrichment analysis involves complex statistical analysis, it requires a computer program. Several tools are available to perform the analysis. One of these computational analysis tools is the web-based Enrichr. Enrichr contains various data sets, such as pathways and protein interactions, gene ontologies, and gene expression in different tissues and cells.

The pathway enrichment analysis and gene ontology analysis were analyzed using the Enrichr online server (<http://maayan-lab.cloud/Enrichr>) (Chen et al., 2013; Kuleshov et al., 2016; Xie et al., 2021). For pathway enrichment analysis, the KEGG 2021 human database was selected to retrieve pathways (Kanehisa and Goto, 2000; Bindea et al., 2009; Jassal et al., 2020). Gene Ontology (GO) enrichment analysis was performed, including those associated with molecular functions, cellular components, and biological processes (Kanehisa and Goto, 2000). Adjusted p-value was used to exclude the influence of multiple comparisons in p-values. Adjusted $p < 0.05$ was considered statistically significant.

Randomness of chromosomal location

The chromosomal location of the loci associated with susceptibility to gout was extracted from the OMIM database (<https://www.omim.org>). The non-randomness of the chromosomal distribution of these loci was statistically evaluated using the method of Tai et al. (1993). The relative nucleotide length of each chromosomal segment to the whole haploid genome was determined using data from the Ensembl Genome Browser (https://asia.ensembl.org/Homo_sapiens/Location/Genome?db=core). To reduce of false positives (type I statistical error reduction), a $p < 0.001$ was considered statistically significant.

RESULTS

We found extracted data from 22 published GWAS studies in the database (Sulem et al., 2011; Lai et al., 2012; Shin et al., 2012; Köttgen et al., 2013; Li et al., 2015; Matsuo et al., 2016; Nakayama et al., 2017, 2020; Chen et al., 2018; Jing et al., 2018; Lee et al., 2019, 2022; Kawamura et al., 2019; Tin et al., 2019; Backman et al., 2021; Dönertaş et al., 2021; Fitzgerald et al., 2022; Jiang et al., 2021; Sandoval-Plata et al., 2021; Toyoda et al., 2022; Lin et al., 2023; Sumpter et al., 2023). Overall, this database contains 64,806 gout patients and 2,856,174 controls. A total of 245

significant associations were initially extracted. For some genes, more than one genetic polymorphism was investigated. Only protein-coding genes were included in the present analysis. Finally, a total of 110 common polymorphic protein-coding loci associated with gout risk were identified and included in the analysis (Table 1).

The results of the KEGG analysis are shown in Table 2. The associated loci were mainly related to ABC transporters (*CFTR*, *ABCC9*, *ABCC8*, *ABCG1*, and *ABCG2*), endocrine and other factor-regulated calcium reabsorption (*PRKCA*, *ATPIA4*, *BDKRB2*, and *VDR*), and gastric acid secretion (*KCNQ1*, *PRKCA*, *CFTR*, and *ATPIA4*) pathways.

Table 1: Polymorphic genes associated with the risk of gout and their cytogenetic locations

Gene Symbols	Locations	Gene Symbols	Locations	Gene Symbols	Locations	Gene Symbols	Locations
<i>PDZK1</i>	1p13.1	<i>SPP1</i>	4q22.1	<i>RUNX1T1</i>	8q21.3	<i>WNT5B</i>	12p13.33
<i>PTGFRN</i>	1p13.1	<i>ADH1B</i>	4q23	<i>PTPRD</i>	9p24.1-p23	<i>SLC38A1</i>	12q13.11
<i>MUC1</i>	1q22	<i>SPCS3</i>	4q34.2	<i>ABCA1</i>	9q31.1	<i>VDR</i>	12q13.11
<i>THBS3</i>	1q22	<i>CDKN2AIP</i>	4q35.1	<i>MED27</i>	9q34.13	<i>INHBC</i>	12q13.3
<i>TRIM46</i>	1q22	<i>TMEM174</i>	5q13.2	<i>MALRD1</i>	10p12.31	<i>R3HDM2</i>	12q13.3
<i>ATPIA4</i>	1q23.2	<i>DST</i>	6p12.1	<i>ARID5B</i>	10q21.2	<i>CUX2</i>	12q24.11-q24.12
<i>SMYD3</i>	1q44	<i>PPARD</i>	6p12.31	<i>SLC16A9</i>	10q21.2	<i>ACAD10</i>	12q24.12
<i>EVA1A</i>	2p12	<i>RUNX2</i>	6p21.1	<i>GRID1</i>	10q23.1-q23.2	<i>ALDH2</i>	12q24.12
<i>GCKR</i>	2p23.3	<i>PRSS16</i>	6p22.1	<i>GLUD1</i>	10q23.2	<i>NAA25</i>	12q24.13
<i>HAGLR</i>	2q31.1	<i>ZSCAN31</i>	6p22.1	<i>SHLD2</i>	10q23.2	<i>TRAFD1</i>	12q24.13
<i>LRP2</i>	2q31.1	<i>CARMIL1</i>	6p22.2	<i>CYP2C8</i>	10q23.33	<i>MLXIP</i>	12q24.31
<i>FRMD4B</i>	3p14.1	<i>H4C5</i>	6p22.2	<i>FGFR2</i>	10q26.13	<i>PIBF1</i>	13q21.33-q22.1
<i>SFMBT1</i>	3p21.1	<i>RREB1</i>	6p24.3	<i>CYP2E1</i>	10q26.3	<i>TCL6</i>	14q32.13
<i>CACNA2D3</i>	3p21.1-p14.3	<i>SLC22A1</i>	6q25.3	<i>ALX4</i>	11p11.2	<i>BDKRB2</i>	14q32.2
<i>SMARCC1</i>	3p21.31	<i>PDE1C</i>	7p14.3	<i>ABCC8</i>	11p15.1	<i>ALDH1A2</i>	15q21.3
<i>RARB</i>	3p24.2	<i>JAZF1</i>	7p15.2-p15.1	<i>SLC22A18AS</i>	11p15.4	<i>UBE2Q2</i>	15q24.2
<i>ZNF639</i>	3q26.33	<i>TPST1</i>	7q11.21	<i>KCNQ1</i>	11p15.5-p15.4	<i>IDH2</i>	15q26.1
<i>NIPAL1</i>	4p12	<i>MLXIPL</i>	7q11.23	<i>AP5B1</i>	11q13.1	<i>SV2B</i>	15q26.1
<i>PPARGC1A</i>	4p15.2	<i>TBL2</i>	7q11.23	<i>CDC42BPG</i>	11q13.1	<i>ACSM2B</i>	16p12.3
<i>CLNK</i>	4p16.1	<i>HGF</i>	7q21.11	<i>MAP3K11</i>	11q13.1	<i>RBFox1</i>	16p13.3
<i>SLC2A9</i>	4p16.1	<i>CNPY4</i>	7q22.1	<i>NRXN2</i>	11q13.1	<i>BCAS3</i>	17q23.2
<i>WDR1</i>	4p16.1	<i>LHFPL3</i>	7q22.1	<i>OVOL1</i>	11q13.1	<i>PRKCA</i>	17q24.2
<i>FAM53A</i>	4p16.3	<i>SRPK2</i>	7q22.3	<i>RPS6KA4</i>	11q13.1	<i>SLC13A3</i>	20q13.12
<i>PRDM8</i>	4q21.21	<i>CFTR</i>	7q31.2	<i>SLC22A11</i>	11q13.1	<i>GABPA</i>	21q21.3
<i>ABCG2</i>	4q22.1	<i>CNTNAP2</i>	7q35-q36.1	<i>CNIH2</i>	11q13.2	<i>ABCG1</i>	21q22.3
<i>DMP1</i>	4q22.1	<i>ASB10</i>	7q36.1	<i>POLD3</i>	11q13.2	<i>PNPLA3</i>	22q13.31
<i>MEPE</i>	4q22.1	<i>PRKAG2</i>	7q36.1	<i>CNTN5</i>	11q22.1		
<i>PKD2</i>	4q22.1	<i>PXDNL</i>	8q11.2-q11.23	<i>ABCC9</i>	12p12.1		

Table 2: Significant findings of pathways and gene ontologies

Results of enrichments	P-value	Adj. p-value	Genes
Pathways (Based on KEGG 2021 Human)			
ABC transporters	1.650e-7	0.00002855	CFTR, ABCC9, ABCC8, ABCG1, ABCG2, ABCA1
Endocrine and other factor-regulated calcium reabsorption	0.0002061	0.01783	PRKCA, ATP1A4, BDKRB2, VDR
Gastric acid secretion	0.0008196	0.04727	KCNQ1, PRKCA, CFTR, ATP1A4
ECM-receptor interaction	0.001416	0.04811	SV2B, SPP1, DMP1, THBS3
Bille secretion	0.001539	0.04811	SLC22A1, ATP1A4, CFTR, ABCG2
Non-alcoholic fatty liver disease	0.001669	0.04811	MLXIP, MLXIPL, PRKAG2, CYP2E1, MAP3K11
Gene Ontologies			
GO: Biological Process 2023			
Vascular Transport (GO:0010232)	3.695e-7	0.0002333	SLC22A1, ABCG2, LRP2, ATP1A4, ABCC9, SLC38A1, SLC13A3
Transport Across Blood-Brain Barrier (GO:0150104)	4.354e-7	0.0002333	SLC13A3, SLC38A1, SLC2A1, ATP1A4, ABCC9, LRP2, ABCG2
Positive Regulation of Transporter Activity (GO:0032411)	0.00005638	0.01511	PDZK1, PPARGC1A, PPARD
Positive Regulation of Transcription by RNA Polymerase II (GO:0045944)	0.00005356	0.01511	MLXIP, BCAS3, HGF, PKD2, MED27, RUNX2, RPS6KA4, MLXIPL, MUC1, ZNF639, RARB, PPARGC1A, RREB1, FGFR2, GABPA, PPARD
Regulation of Transcription by RNA Polymerase II (GO:0006357)	0.0002302	0.04935	SLC22A1, ABCG2
GO: Cellular Component 2023			
External Side of Apical Plasma Membrane (GO:0098591)	0.0003019	0.02959	ABCG2, SLC38A1

The gene ontology (GO) analysis consisted of three functional parts, including biological process (BP), cellular component (CC), and molecular function (MF). The top four biological processes were vascular transport (GO: 0010232), transport across blood-brain barrier (GO: 0150104), positive regulation of transporter activity (GO: 0032411), and positive regulation of transcription by RNA polymerase II (GO: 0045944). The top cellular component was the external side of the apical plasma membrane (GO: 0098591) with two *ABCG2*, and *SLC38A1* genes. There was no statistically

significant gene ontology analysis for the molecular functions (Table 2).

Shared polymorphic loci between gout and selected traits, based on GWAS Catalog 2023 was investigated. The results were summarized in Table 3. Chronic kidney disease, kidney stones, type 2 diabetes, fasting glucose, triglyceride levels, metabolic syndrome, coronary artery disease, diastolic and systolic blood pressure, resistant hypertension, systemic lupus erythematosus, alcohol dependence, schizophrenia, rate of cognitive decline in Alzheimer's disease, Alzheimer's disease, bipolar disorder or major depressive disorder,

COVID-19 (critical illness vs population or mild symptoms), severe COVID-19 infection, COVID-19 (hospitalized vs population) were selected traits which had shared polymorphic loci with gout.

Of 110 potentially gout-associated polymorphic loci, 3 (*MUC1*, *THBS3*, and *TRIM46*), 3 (*CLNK*, *SLC2A9*, and *WDR1*), 5 (*ABCG2*, *DMP1*, *MEPE*, *PKD2*, and *SPP1*), 4 (*PRSS16*, *ZSCAN31*, *CARMIL1*, and *H4C5*), 8 (*CDC42BPG*, *MAP3K11*, *NRXN2*, *OVOL1*, *RPS6KA4*, *SLC22A11*, *CNIH2*, and

POLD3), 4 (*SLC38A1*, *VDR*, *INHBC*, and *R3HDM2*), and 5 (*CUX2*, *ACAD10*, *ALDH2*, *NAA25*, and *TRAFDI*) genes were located on the human 1q22, 4p16.1, 6p21.1-p21.2, 11q13.1-q13.2, 12q13.11-q13.3, and 12q24.1 chromosome segments, respectively. These chromosomal distributions are not random (Table 4). There was no statistical evidence that the other gout-associated loci non-randomly distributed on the chromosomes.

Table 3: Shared polymorphic loci between gout and selected traits

Traits	P-value	Adjusted p-value	Odds Ratio	Combined score
Chronic Kidney Disease	7.053e-13	1.303e-10	31.60	884.09
Kidney Stones	0.0001511	0.001402	34.50	303.53
Type 2 Diabetes	1.310e-7	0.000006365	4.68	74.09
Fasting Glucose	0.0007888	0.004411	7.47	53.37
Triglyceride Levels	1.458e-9	1.282e-7	5.93	120.65
Metabolic Syndrome	0.0001108	0.001106	11.68	106.38
Coronary Artery Disease	0.000004885	0.0001248	5.20	63.56
Coronary Heart Disease	0.0005263	0.003125	11.76	88.82
Diastolic Blood Pressure	0.00003741	0.0004866	3.79	38.61
Systolic Blood Pressure	0.0002812	0.002097	3.00	24.53
Resistant Hypertension	0.0001770	0.001550	32.47	280.52
Cardiovascular Disease Risk Factors	5.255e-8	0.000003235	66.96	1122.42
Systemic Lupus Erythematosus	0.0003676	0.002425	4.97	39.31
Alcohol Dependence	0.001416	0.006953	14.90	97.77
Schizophrenia	0.0001082	0.001081	3.28	29.99
Rate of Cognitive Decline in Alzheimer's Disease	0.0008905	0.004754	10.15	71.27
Alzheimer's Disease or HDL Levels (Pleiotropy)	0.002288	0.009823	33.16	201.60
Anorexia Nervosa, Attention-Deficit/Hyperactivity Disorder, Autism Spectrum Disorder, Bipolar Disorder, Major Depression, Obsessive-Compulsive Disorder, Schizophrenia, or Tourette Syndrome (Pleiotropy)	0.002017	0.009130	8.04	49.92
Bipolar Disorder or Major Depressive Disorder	0.006418	0.02144	8.47	42.77
Bipolar Disorder and Schizophrenia	0.007807	0.02517	7.86	38.16
COVID-19 (Critical Illness vs Population or Mild Symptoms)	0.007157	0.02331	17.36	85.75
Severe COVID-19 Infection	0.008444	0.02698	3.21	15.34
COVID-19 (Hospitalized vs Population)	0.01420	0.04236	6.25	26.59

Table 4: Non-random distribution of gout susceptible loci on human chromosomes

Chromosome bands	Number of loci	F	df	P-value
1q22	3	57.38	216, 6	0.000024
4p16.1	3	16.21	216, 6	0.000942
4q22.1	5	25.16	212, 10	<0.00001
6p22.1-p22.2	4	21.62	214, 8	0.000044
11q13.1-q13.2	8	46.67	206, 16	<0.00001
12q13.11-q13.3	4	10.49	214, 8	0.000680
12q24.1	5	30.10	212, 10	<0.00001

DISCUSSION

This study used data available in the GWAS database on polymorphic loci associated with gout risk. A total of 110 common polymorphic protein-coding loci associated with gout risk were identified and included in the analysis. Enrichment analysis was then performed using the Enrichr tool. The present study showed that in the gene ontology analysis, BP mainly focuses on vascular transport, transport across the blood-brain barrier, positive regulation of transporter activity, and positive regulation of transcription by RNA polymerase II, CC mainly focuses on the external side of the apical plasma membrane.

Previously, Qiu and colleagues reported differentially expressed genes (DEGs) in gout using the GEO database (Qiu et al., 2022). They reported that the results of gene ontology analysis of the DEGs were mainly enriched in immune and inflammatory response, cytokine and growth factor activities; also KEGG pathway analysis showed that the DEGs were mainly related to chemokine signalling pathway and cytokine-cytokine receptor interaction (Qiu et al., 2022). It should be noted that the present results are not only not similar to those of Qiu et al., but also quite different. There is no commonality in the results of the enrichment analysis between the Qiu study and the present study. Among the polymorphic genes associated with gout, there is no gene involved in the immune system. At present, it is very difficult to interpret this discrepancy. However, some suggestions

can be made. First, they used only one data set, whereas we used all available data sets. Second, the analysis of differentially expressed genes (DEGs) was based on a very small sample size (12 participants including 6 gout patients and 6 healthy controls), whereas the GWAS data were obtained from very large samples. Third, the subjects in Qiu's study were all Chinese males, whereas the present study used data from both sexes belonging to different ethnic groups. Finally, the differentially expressed genes and the polymorphic genes are two different sets of genes that are involved in the pathogenesis of gout.

As mentioned in the introduction, gout is often associated with other conditions such as hypertension, cardiovascular disease, diabetes, dyslipidemia, chronic kidney disease, and kidney stones (Dalbeth et al., 2021). The present study showed that these traits shared polymorphic loci with gout (Table 3). Surprisingly, both susceptibility to COVID-19 and mortality due to COVID-19 shared polymorphic protein-coding genes with gout (Table 3). It should be noted that there are significant associations between gout and both susceptibility to COVID-19 and COVID-19-related death (Dalbeth and Robinson, 2021; Peng et al., 2022; Nissen et al., 2022; Topless et al., 2022). An association between low serum urate concentrations and the risk of neurodegenerative diseases, such as Parkinson's disease and Alzheimer's disease, has been reported previously (Li et al., 2017; Singh and Cleveland, 2019). A meta-analysis of four co-

hort studies reported that gout and hyperuricemia might reduce the risk of AD (Pan et al., 2021). Interestingly, Alzheimer's disease shared polymorphic loci with gout (Table 3). Previously, it has been reported that genes associated with the risk of Alzheimer's disease are not randomly distributed on human chromosomes. One of the human chromosomal segments carrying Alzheimer's disease-associated genes is 6p21 (Saadat, 2016). Interestingly, the present study indicated that 6p221-p22.2 chromosome segment which is located in the vicinity of 6p21 and obviously had linkage disequilibrium with each other, carries gout associated loci (Table 4).

The present finding of non-random chromosomal distribution of gout-associated loci is similar to the non-random distribution of some other disease-associated (such as breast and gastric cancers, Alzheimer's disease) genes on human chromosomes, which supports non-random distribution of genes in the construction of human chromosomes (Saify and Saadat, 2012; Saadat, 2014, 2016; Mahjoub and Saadat, 2018).

The present findings suggest the possibility of designing and developing a laboratory diagnostic test method using the genetic variations on the human chromosome segments 1q22, 4p16.1, 6p21.1-p21.2, 11q13.1-q13.2, 12q13.11-q13.3, and 12q24.1 for use in mass screening programs to identify individuals at high risk for developing gout.

Conflict of interest

The author declares no conflict of interest.

REFERENCES

- Backman JD, Li AH, Macketta A, Sun D, Mbatchou J, Kessler MD, et al. Exome sequencing and analysis of 454,787 UK Biobank participants. *Nature*. 2021;599(7886):628-34.
- Bindea G, Mlecnik B, Hackl H, Charoentong P, Tosolini M, Kirilovsky A, et al. ClueGO: a Cytoscape plugin to decipher functionally grouped gene ontology and pathway annotation networks. *Bioinformatics*. 2009; 25:1091-3.
- Bleyer AJ, Hart TC. Genetic factors associated with gout and hyperuricemia. *Adv Chronic Kidney Dis*. 2006;13:124-30.
- Chen CJ, Tseng CC, Yen JH, Chang JG, Chou WC, Chu HW, et al. ABCG2 contributes to the development of gout and hyperuricemia in a genome-wide association study. *Sci Rep*. 2018;8(1):3137.
- Chen EY, Tan CM, Kou Y, Duan Q, Wang Z, Meirelles GV, et al. Enrichr: interactive and collaborative HTML5 gene list enrichment analysis tool. *BMC Bioinformatics*. 2013;14:128.
- Dalbeth N, Robinson PC. Patients with gout: an under-recognised group at high risk of COVID-19. *Lancet Rheumatol*. 2021;3(5):e317-8.
- Dalbeth N, Gosling AL, Gaffo A, Abhishek A. Gout. *Lancet*. 2021;397(10287):1843-55.
- Dönertaş HM, Fabian DK, Valenzuela MF, Partridge L, Thornton JM. Common genetic associations between age-related diseases. *Nat Aging*. 2021;1(4):400-12.
- Dong Z, Guo S, Yang Y, Wu J, Guan M, Zou H, et al. Association between ABCG2 Q141K polymorphism and gout risk affected by ethnicity and gender: a systematic review and meta-analysis. *Int J Rheum Dis*. 2015;18:382-91.
- Emmerson BT, Nagel SL, Duffy DL, Martin NG. Genetic control of the renal clearance of urate: a study of twins. *Ann Rheum Dis*. 1992;51:375-7.
- Fitzgerald T, Birney E. CNest: A novel copy number association discovery method uncovers 862 new associations from 200,629 whole-exome sequence datasets in the UK Biobank. *Cell Genom*. 2022;2(8):100167.
- Jassal B, Matthews L, Viteri G, Gong C, Lorente P, Fabregat A, et al. The reactome pathway knowledge-base. *Nucleic Acids Res*. 2020;48(D1):D498-D503.
- Jiang L, Zheng Z, Fang H, Yang J. A generalized linear mixed model association tool for biobank-scale data. *Nat Genet*. 2021;53:1616-21.
- Jing J, Ekici AB, Sitter T, Eckardt KU, Schaeffner E, Li Y, et al. Genetics of serum urate concentrations and gout in a high-risk population, patients with chronic kidney disease. *Sci Rep*. 2018;8(1):13184.
- Kanehisa M, Goto S. KEGG: kyoto encyclopedia of genes and genomes. *Nucleic Acids Res*. 2000; 28(1): 27-30.

- Kawaguchi M, Nakayama A, Aoyagi Y, Nakamura T, Shimizu S, Kawamura Y, et al. Both variants of AICF and BAZ1B genes are associated with gout susceptibility: a replication study and meta-analysis in a Japanese population. *Hum Cell*. 2021;34:293-9.
- Kawamura Y, Nakaoka H, Nakayama A, Okada Y, Yamamoto K, Higashino T, et al. Genome-wide association study revealed novel loci which aggravate asymptomatic hyperuricaemia into gout. *Ann Rheum Dis*. 2019;78:1430-7.
- Köttgen A, Albrecht E, Teumer A, Vitart V, Krumsiek J, Hundertmark C, et al. Genome-wide association analyses identify 18 new loci associated with serum urate concentrations. *Nat Genet*. 2013;45:145-54.
- Krishnan E, Lessov-Schlaggar CN, Krasnow RE, Swan GE. Nature versus nurture in gout: a twin study. *Am J Med*. 2012;125:499-504.
- Kuleshov MV, Jones MR, Rouillard AD, Fernandez NF, Duan Q, Wang Z, et al. Enrichr: a comprehensive gene set enrichment analysis web server 2016 update. *Nucleic Acids Res*. 2016; 44(W1):W90-7.
- Kuo CF, Grainge MJ, See LC, Yu KH, Luo SF, Valdes AM, et al. Familial aggregation of gout and relative genetic and environmental contributions: a nationwide population study in Taiwan. *Ann Rheum Dis*. 2015;74:369-74.
- Lai HM, Chen CJ, Su BY, Chen YC, Yu SF, Yen JH, et al. Gout and type 2 diabetes have a mutual inter-dependent effect on genetic risk factors and higher incidences. *Rheumatology (Oxford)*. 2012;51:715-20.
- Lee CJ, Chen TH, Lim AMW, Chang CC, Sie JJ, Chen PL, et al. Phenome-wide analysis of Taiwan Biobank reveals novel glycemia-related loci and genetic risks for diabetes. *Commun Biol*. 2022;5(1):1175.
- Lee MG, Hsu TC, Chen SC, Lee YC, Kuo PH, Yang JH, et al. Integrative genome-wide association studies of eQTL and GWAS data for gout disease susceptibility. *Sci Rep*. 2019;9(1):4981.
- Lee YH, Seo YH, Kim JH, Choi SJ, Ji JD, Song GG. Associations between *SLC2A9* polymorphisms and gout susceptibility: A meta-analysis. *Z Rheumatol*. 2017;76:64-70.
- Li C, Li Z, Liu S, Wang C, Han L, Cui L, et al. Genome-wide association analysis identifies three new risk loci for gout arthritis in Han Chinese. *Nat Commun*. 2015;6:7041.
- Li X, Meng X, Timofeeva M, Tzoulaki I, Tsilidis KK, Ioannidis JP, et al. Serum uric acid levels and multiple health outcomes: umbrella review of evidence from observational studies, randomised controlled trials, and Mendelian randomisation studies. *BMJ*. 2017;357:j2376.
- Lin CY, Chang YS, Liu TY, Huang CM, Chung CC, Chen YC, et al. Genetic contributions to female gout and hyperuricaemia using genome-wide association study and polygenic risk score analyses. *Rheumatology (Oxford)*. 2023;62:638-46.
- Mahjoub G, Saadat M. Non-random distribution of gastric cancer susceptible loci on human chromosomes. *EXCLI J*. 2018;17:802-7.
- Matsuo H, Yamamoto K, Nakaoka H, Nakayama A, Sakiyama M, Chiba T, et al. Genome-wide association study of clinically defined gout identifies multiple risk loci and its association with clinical subtypes. *Ann Rheum Dis*. 2016;75 652-9.
- Nakayama A, Nakaoka H, Yamamoto K, Sakiyama M, Shaukat A, Toyoda Y, et al. GWAS of clinically defined gout and subtypes identifies multiple susceptibility loci that include urate transporter genes. *Ann Rheum Dis*. 2017;76:869-77.
- Nakayama A, Nakatochi M, Kawamura Y, Yamamoto K, Nakaoka H, Shimizu S, et al. Subtype-specific gout susceptibility loci and enrichment of selection pressure on *ABCG2* and *ALDH2* identified by subtype genome-wide meta-analyses of clinically defined gout patients. *Ann Rheum Dis*. 2020;79:657-65.
- Nissen CB, Hendricks O, Schreiber K. Women with gout and COVID-19-an unfortunate combination? *Lancet Rheumatol*. 2022;4:e233-4.
- Pan SY, Cheng RJ, Xia ZJ, Zhang QP, Liu Y. Risk of dementia in gout and hyperuricaemia: a meta-analysis of cohort studies. *BMJ Open*. 2021;11(6):e041680.
- Peng H, Wu X, Xiong S, Li C, Zhong R, He J, et al. Gout and susceptibility and severity of COVID-19: A bidirectional Mendelian randomization analysis. *J Infect*. 2022;85(3):e59-e61.
- Qiu K, Zeng T, Liao Y, Min J, Zhang N, Peng M, et al. Identification of inflammation-related biomarker ProADM for male patients with gout by comprehensive analysis. *Front Immunol*. 2022;12:798719.
- Saadat M. Distributions of susceptibility loci of Parkinson's disease and multiple sclerosis on human chromosomes. *EXCLI J*. 2014;13:724-7.
- Saadat M. Distributions of susceptibility loci to late onset Alzheimer's disease on human chromosomes. *EXCLI J*. 2016;15:403-5.

- Saify K, Saadat M. Non-random distribution of breast cancer susceptibility loci on human chromosomes. *Breast Cancer Res Treat.* 2012;136:315-8.
- Sandoval-Plata G, Morgan K, Abhishek A. Variants in urate transporters, ADH1B, GCKR and MEPE genes associate with transition from asymptomatic hyperuricaemia to gout: results of the first gout versus asymptomatic hyperuricaemia GWAS in Caucasians using data from the UK Biobank. *Ann Rheum Dis.* 2021;80:1220-6.
- Shin J, Kim Y, Kong M, Lee C. Genetic architecture for susceptibility to gout in the KARE cohort study. *J Hum Genet.* 2012;57:379-84.
- Singh JA, Cleveland JD. Gout and the risk of Parkinson's disease in older adults: a study of US Medicare data. *BMC Neurol.* 2019;19(1):4.
- Subramanian A, Tamayo P, Mootha VK, Mukherjee S, Ebert BL, Gillette MA, et al. Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proc Natl Acad Sci U S A.* 2005;102:15545-50.
- Sulem P, Gudbjartsson DF, Walters GB, Helgadóttir HT, Helgason A, Gudjonsson SA, et al. Identification of low-frequency variants associated with gout and serum uric acid levels. *Nat Genet.* 2011;43:1127-30.
- Sumpter NA, Takei R, Cadzow M, Topless RKG, Phipps-Green AJ, Murphy R, et al. Association of gout polygenic risk score with age at disease onset and tophaceous disease in European and Polynesian men with gout. *Arthritis Rheumatol.* 2023;75:816-25.
- Tai JJ, Hou CD, Wang-Wuu S, Wang CH, Leu SY, Wu KD. A method for testing the non-randomness of chromosomal breakpoints. *Cytogenet Cell Genet.* 1993;63:147-50.
- Tin A, Marten J, Halperin Kuhns VL, Li Y, Wuttke M, Kirsten H, et al. Target genes, variants, tissues and transcriptional pathways influencing human serum urate levels. *Nat Genet.* 2019;51:1459-74.
- Topless RK, Gaffo A, Stamp LK, Robinson PC, Dalbeth N, Merriman TR. Gout and the risk of COVID-19 diagnosis and death in the UK Biobank: a population-based study. *Lancet Rheumatol.* 2022;4:e274-81.
- Toyoda Y, Nakayama A, Nakatochi M, Kawamura Y, Nakaoka H, Yamamoto K, et al. Genome-wide meta-analysis between renal overload type and renal underexcretion type of clinically defined gout in Japanese populations. *Mol Genet Metab.* 2022;136:186-9.
- Voruganti VS, Göring HH, Mottl A, Franceschini N, Haack K, Laston S, et al. Genetic influence on variation in serum uric acid in American Indians: the strong heart family study. *Hum Genet.* 2009;126:667-76.
- Wilk JB, Djousse L, Borecki I, Atwood LD, Hunt SC, Rich SS, et al. Segregation analysis of serum uric acid in the NHLBI Family Heart Study. *Hum Genet.* 2000;106:355-9.
- Xie Z, Bailey A, Kuleshov MV, Clarke DJB, Evangelista JE, Jenkins SL, et al. Gene set knowledge discovery with Enrichr. *Curr Protoc.* 2021;1(3):e90.
- Zou Y, Du J, Zhu Y, Xie X, Chen J, Ling G. Associations between the SLC22A12 gene and gout susceptibility: a meta-analysis. *Clin Exp Rheumatol.* 2018;36:442-7.
-