

Supplementary information to:

Original article:

BIOENGINEERING SCALABLE AND DRUG-RESPONSIVE *IN VITRO* HUMAN MULTICELLULAR NON-ALCOHOLIC FATTY LIVER DISEASE MICROTISSUES ENCAPSULATED IN THE LIVER EXTRACELLULAR MATRIX-DERIVED HYDROGEL

Negar Asadollahi^{1,8} , Mohammad Amin Hajari² , Mahmoud Alipour Choshali³ ,
Mohammad Ajoudanian⁴ , Seyed Ali Ziai⁵ , Massoud Vosough^{1,6} *, Abbas Piryaee^{7*} 

- ¹ Department of Regenerative Medicine, Cell Science Research Center, Royan Institute for Stem Cell Biology and Technology, ACECR, Tehran, Iran
- ² Department of Cell Engineering, Cell Science Research Center, Royan Institute for Stem Cell Biology and Technology, ACECR, Tehran, Iran
- ³ Department of Stem Cells and Developmental Biology, Cell Science Research Center, Royan Institute for Stem Cell Biology and Technology, ACECR, Tehran, Iran
- ⁴ Department of Medical Biotechnology, School of Advanced Technologies in Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran
- ⁵ Department of Pharmacology, School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran
- ⁶ Experimental Cancer Medicine, Institution for Laboratory Medicine, Karolinska Institute, Huddinge, Sweden
- ⁷ Department of Biology and Anatomical Sciences, School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran
- ⁸ Department of Developmental Biology, University of Science and Culture, Tehran, Iran

* **Corresponding authors:** Abbas Piryaee, Department of Biology and Anatomical Sciences, School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran.

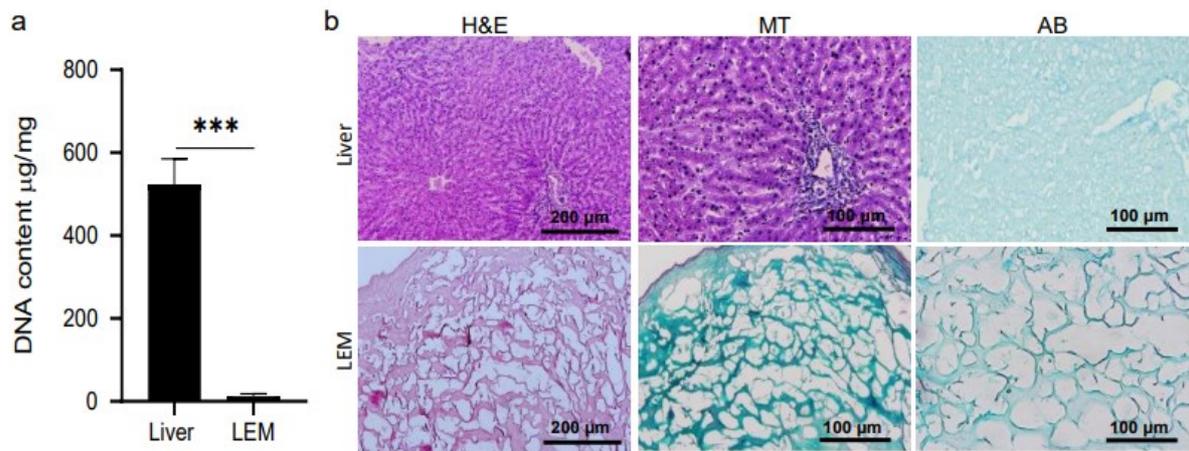
Tel: +989126971784, E-mail: piryae@sbmu.ac.ir

Massoud Vosough, Department of Regenerative Medicine, Cell Science Research Center, Royan Institute for Stem Cell Biology and Technology, ACECR, Tehran, Iran.

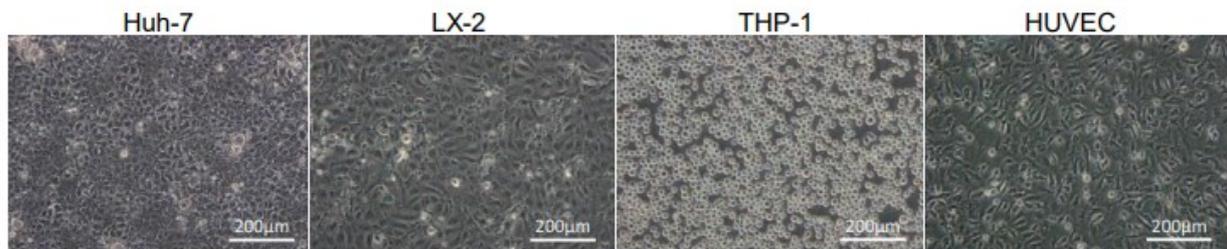
Tel: +989121196454, E-mail: masvos@royaninstitute.org

<https://dx.doi.org/10.17179/excli2023-6878>

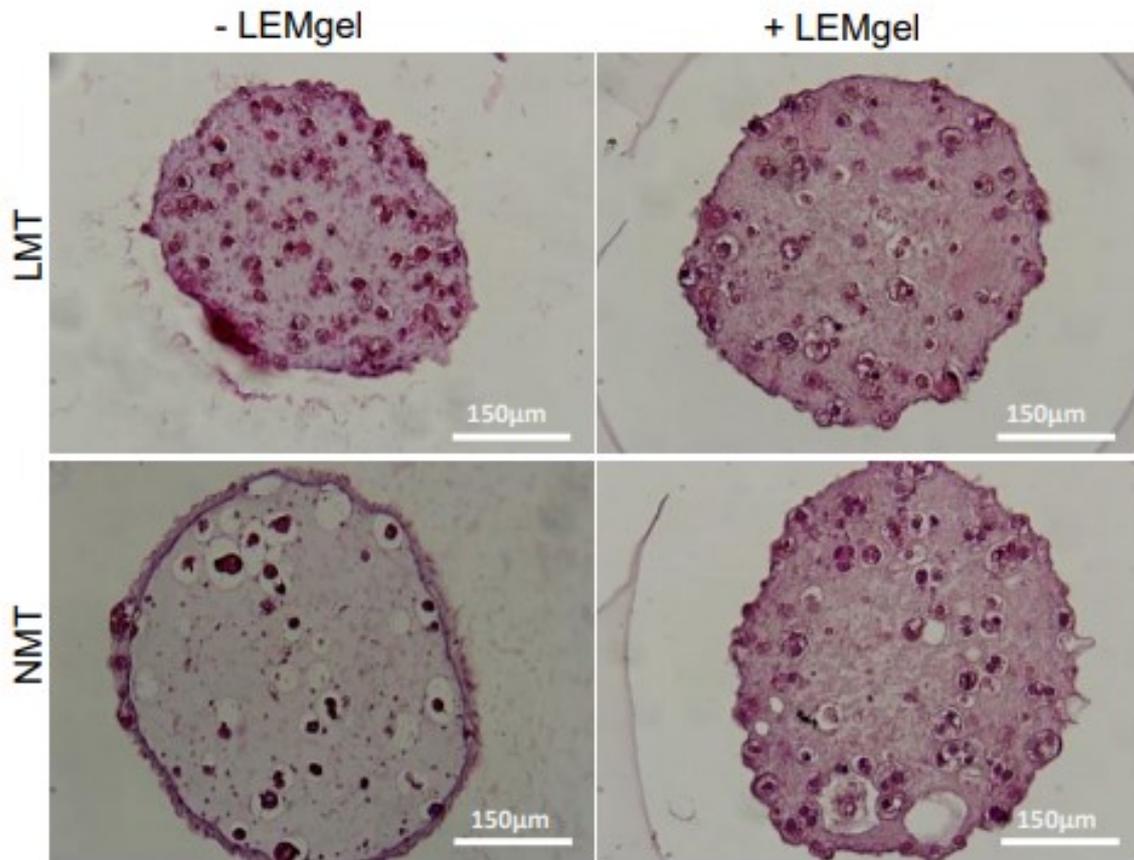
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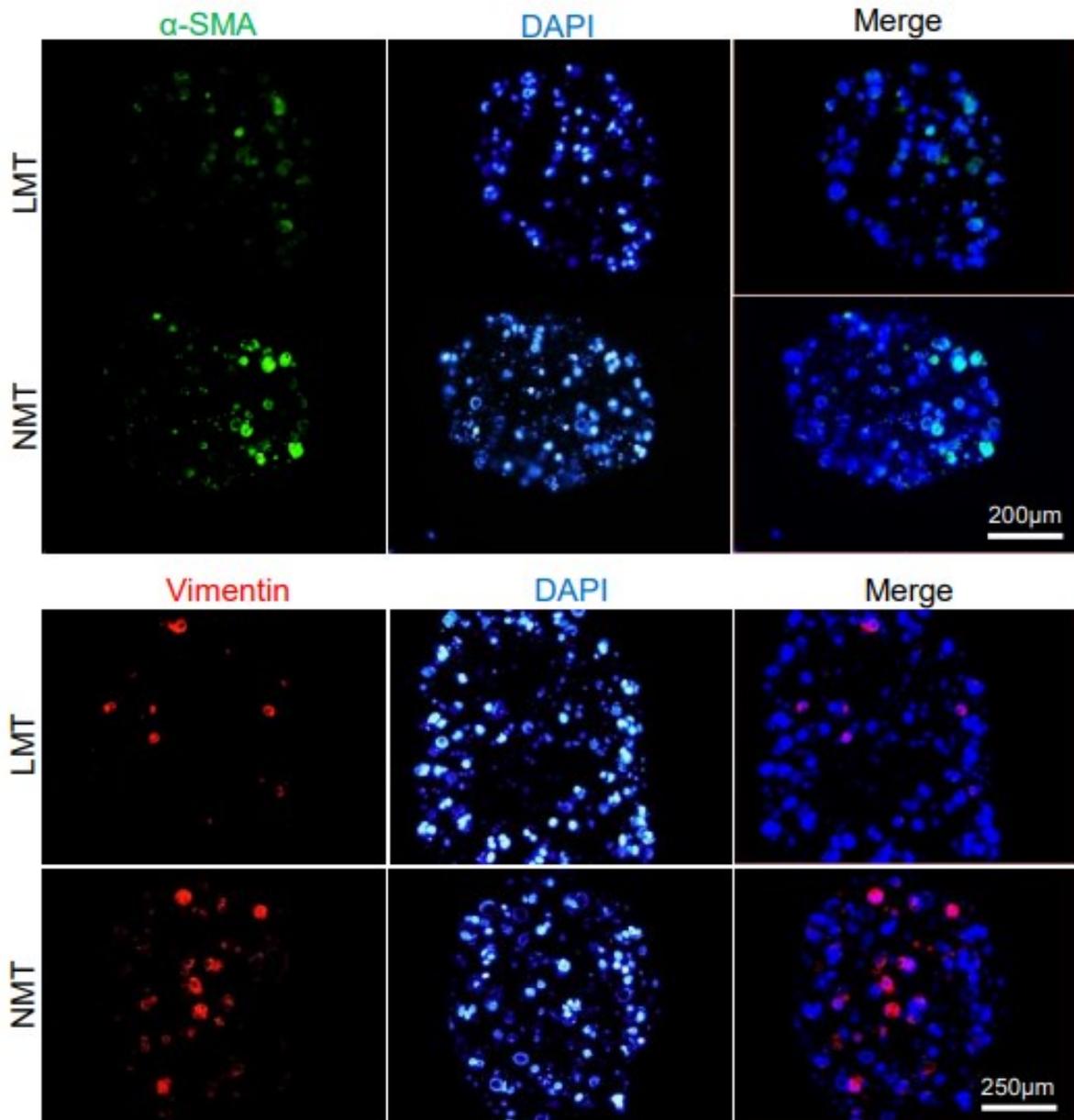
Supplementary Figure 1: Characterization of the decellularized liver-derived extracellular matrix (LEM). **a)** Quantification of DNA content in the liver and LEM. **b)** Histological evaluations of the liver and the resultant LEM. Hematoxylin and eosin (H&E) staining showed that, compared to the native liver, there are no nuclei in the LEM. Preservation of collagen and glycosaminoglycans (GAGs) in the LEM was demonstrated using Masson's trichrome (MT) and Alcian blue (AB) staining, respectively.



Supplementary Figure 2: Phase-contrast micrographs of Huh7, LX-2, and THP-1 cell lines and HUVEC, demonstrated the cells morphology in two-dimensional culture condition.



Supplementary Figure 3: Light micrographs from Hematoxylin and eosin-stained sections of the LMTs and NMTs generated in the presence or absence of LEMgel on day 8. As it is visible, the overall structures of the +LEMgel microtissues are more appropriate with homogenous distribution of live cells. While the -LEMgel microtissues revealed visible destruction in their structure and there are many necrotic cells with pyknotic nuclei, particularly in the NMT.



Supplementary Figure 4: Representative immunofluorescence micrographs from the microtissue sections stained against Vimentin and α -SMA on day 8. The micrographs revealed that there are more Vimentin-positive and α -SMA-positive cells in the NMTs in comparison to the LMTs.

Supplementary Table 1: Primer sequences used in qRT-PCR

Genes	Sequence
<i>CPT1</i>	F: ATCAATCGGACTCTGGAAACGG R: TCAGGGAGTAGCGCATGGT
<i>CD36</i>	F: CTTTGGCTTAATGAGACTGGGAC R: GCAACAAACATCACCACACCA
<i>SREBP</i>	F: CGGAACCATCTTGGCAACAGT R: CGCTTCTCAATGGCGTTGT
<i>CYP2E1</i>	F: GTGATGCACGGCTACAAGG R: GGGTGGTCAGGGAAAACCG
<i>IL 6</i>	F: AGGAGACTTGCCTGGTGAAA R: CAGGGGTGGTTATTGCATCT
<i>IL 8</i>	F: TAGCAAATTGAGGCCAAGG R: AGCAGACTAGGGTTGCCAGA
<i>IL 10</i>	F: AAGCTGAGAACCAAGACCCA R: AAGGCATTCTTCACCTGCTC
<i>TGF-β</i>	F: GAAACCCACAACGAAATCTATGA R: TAACTTGAGCCTCAGCAGAC
<i>TNF-α</i>	F: CCTCTCTAATCAGCCCTCTG R: GAGGACCTGGGAGTAGATGAG
<i>MMP-2</i>	F: ATTGTATTTGATGGCATCGCTC R: ATTCATTCCCTGCAAAGAACAC
<i>COL1A1</i>	F: ATGCCTGGTGAACGTGGT R: AGGAGAGCCATCAGCACCT
<i>PCK1</i>	F: GGCTGAAGAAGTATGACAACTG R: AAATCCTCCTCTGACATCCA
<i>G6P</i>	F: AACATCGCCTGCGTTATCCTC R: ACGTCCCGGATGATCCCAA
<i>GAPDH</i>	F: GAAATCCCATCACCATCTTCC R: GGCTGTTGTCATACTTCTCAT