

## Original article:

# COMPARATIVE GROWTH ANALYSIS OF CAPSULATED (VI+) AND ACAPSULATED (VI-) *SALMONELLA* TYPHI ISOLATES IN HUMAN BLOOD

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## ABSTRACT

*Salmonella enterica* serovar Typhi (*S. Typhi*) is a human restricted pathogen. It biosynthesizes a virulence capsular polysaccharide named as Vi antigen. *S. Typhi* regulates expression of genes involved in the biosynthesis of Vi antigen in response to osmolarity. Beside Vi-positive isolates, Vi-negative (acapsulated) isolates are also pathogenic. However, Vi-positive isolates are more prevalent. The present study was planned to investigate comparative growth of Vi-positive and Vi-negative *S. Typhi* isolates in an *ex vivo* human whole blood model. Four isolates of each type were tested for growth in human whole blood and in an enrichment medium (Tryptic soy broth-TSB) as a control. It was found that capsulated (Vi-positive) strains formed smooth circular colonies and grew with shorter lag and generation time than Vi-negative isolates. Overall growth pattern of *S. Typhi* isolates both *in vitro* and *ex vivo* conditions showed that Vi-positive isolates grew at a faster rate. Especially in human blood, the lag time of acapsulated isolates was almost doubled as compared to capsulated *S. Typhi* isolates. It was also observed that Vi-negative isolates reduced in number up to 81 % during the first 12 hours of incubation in human whole blood. Interestingly, both types of isolates had similar growth curve in TSB indicating that Vi capsule is dispensable for bacterial growth *in vitro*. This study shows for the first time that absence of capsular antigen retards the growth of Vi-negative isolates on initial contact with human blood, but with passage of time they adjust themselves according to the new environment.

**Keywords:** *Salmonella* Typhi, Vi+, Vi-, growth, human blood

## INTRODUCTION

*Salmonella enterica* serovar Typhi is an exclusively human pathogen and a leading cause of enteric fever worldwide. It causes an acute, systemic infection of the reticulo-endothelial system. Our knowledge of the actual mechanism of pathogenesis of *S. Typhi* is limited because it infects only humans, which means that animal models are not available to study host-pathogen interactions. Most of the current understandings about the pathogenicity of *S. Typhi* have

been extrapolated from *S. Typhimurium* infections in mice. A major limitation for using this murine model is that *S. Typhimurium* does not cause typhoid fever in humans but instead causes a localized gastroenteritis resulting in diarrhea. *S. Typhi* produces a virulence factor, the Vi capsular antigen which is absent in *S. Typhimurium*. Vi capsule is synthesized by genes located on a large pathogenicity island designated as SPI-7 which is not present in the genome of *S. Typhimurium*. SPI-7 is genetically unstable (Bueno et

al., 2004; Nair et al., 2004) and its deletion may give rise to acapsulated *S. Typhi* isolates. Therefore, on the basis of capsule formation, *S. Typhi* can be grouped into two types: capsulated (Vi-positive) and acapsulated (Vi-negative). Both types have been identified and isolated from typhoid patient's blood (Baker et al., 2005; Pulickal et al., 2013; Wain et al., 2005) and volunteer studies have shown that both can cause enteric fever (Hone et al., 1988).

During the course of infection, Vi capsular antigen protects bacteria against host immune system due to its anti-inflammatory (Crawford et al., 2013; Haneda et al., 2009; Raffatellu et al., 2007), antiopsonic and antiphagocytic property (Wilson et al., 2011). The expression of genes involved in the Vi capsular biosynthesis is greatly influenced by the environmental conditions. The osmolarity of the lumen of small intestine is equivalent to 0.3 M NaCl (Sleator et al., 2007) and bacterial exposure to this high osmolarity results in drastic decrease in the expression of Vi antigen (Tartera and Metcalf, 1993; Tran et al., 2010). It is expressed when *S. Typhi* transits from intestinal lumen into the ileal mucosa under low osmolarity environment which may explain the protective role of capsule during systemic infection.

Role of Vi antigen in survival of *S. Typhi* has been studied using macrophage (Size-more et al., 1997; Wilson et al., 2011), neutrophil (Wangdi et al., 2014) and epithelial cell lines. The role of Vi antigen in serum resistance has also been reported (Bravo et al., 2008; Looney and Steigbigel, 1986). However, during infection, the behavior of capsular and acapsular variants of *S. Typhi* in whole human blood is still in question. Two conflicting reports have described the role of Vi antigen in protection of *S. Typhi* from inhibitory effect of human serum (Bravo et al., 2008; Looney and Steigbigel, 1986). According to Looney and Steigbigel (1986), Vi-positive strains are more resistant to lysis by non-immune serum than Vi-negative isolates, whereas Bravo et al.

(2008) demonstrated that the Vi antigen does not contribute to serum resistance.

This study is based on the comparative growth behavior of Vi-negative and Vi-positive variants of *S. Typhi* in human whole blood as an *ex vivo* model. This comparative study on survival and growth of naturally occurring Vi-negative and Vi-positive *S. Typhi* strains was conducted to investigate whether Vi antigen expression is enhanced within human blood due to low osmolarity conditions, and consequently there are different growth rates of Vi-negative and Vi-positive *S. Typhi* isolates in blood.

## MATERIALS AND METHODS

### *Bacterial strains*

Four Vi-positive and four Vi-negative *S. Typhi* isolates cultured from blood samples of typhoid patients were investigated in this study. Blood samples were collected from different patients suffering from typhoid fever in various hospitals in Faisalabad region of Pakistan. All isolates were revived from National Institute for Biotechnology and Genetic Engineering (NIBGE) stocks stored in 20 % glycerol at -20 °C (Baker et al., 2005).

### *In vitro growth analysis*

Tryptic soy broth (TSB), a universal culture medium free from inhibitors and indicators with osmolarity (300 mosmol/liter) similar to human blood (Scharfman et al., 1996), was used for reviving the *S. Typhi* stock cultures for in vitro growth analysis. Growth in TSB was subcultured on MacConkey agar. A single colony was reinoculated in 3 ml TSB and cultured at 37 °C with shaking at 200 rpm for at least 18 hours. Fifty µl of this overnight culture were inoculated into 3 ml of fresh TSB and cultured at 37 °C with shaking at 200 rpm. At time points of 0, 2, 4, 6, 8, 10, 12, 14, 16, 18 and 24 hours after inoculation, samples of bacterial cultures were collected and used for analysis after making ten fold serial dilution in autoclaved distilled water. The CFU were quantified by plating the appropriate dilutions on Mac-

Conkey agar. Each sample was analyzed in triplicate and the analysis was repeated at least three times. The bacterial counts were transformed into logarithms and their average values of CFU/ml were used to plot the growth curve. The generation time was determined by comparing bacterial cell numbers at two time points of the exponential growth phase using following formula (Liu et al., 2006):

$$\text{Generation time} = 0.301 (T_2 - T_1) / (\log N - \log N_0)$$

$T_1$  is the first time point (minutes) and  $T_2$  is the second time point of the exponential phase,  $N_0$  is the number of bacterial cells at  $T_1$ , and  $N$  is the number of bacterial cells at  $T_2$ .

#### **Human whole blood as ex vivo growth model**

Human whole blood has been used as an *ex vivo* model for studying the human restricted pathogens including *Neisseria meningitidis*, *Listeria monocytogenes*, and group A and group B *Streptococcus* species (Echenique-Rivera et al., 2011; Graham et al., 2005; Mereghetti et al., 2008; Toledo-Arana et al., 2009). *S. Typhi* is also a human restricted pathogen. Therefore, in this study human blood from healthy volunteers was used as *ex vivo* model for comparative growth analysis of Vi-negative and Vi-positive *S. Typhi* isolates with that of TSB medium (as *in vitro* model). Human venous blood was collected (in heparin) from 4 healthy volunteers (two males and two females, ages ranged from 25 to 35 years) who were not on any antimicrobial treatment for the past eight months before this study.

Bacterial loads in typhoid patients can reach up to  $10^4$  bacteria/ml (Massi et al., 2005; Wain et al., 2001). Therefore, approximately  $10^3$  bacteria from early exponential phase culture within TSB were mixed with heparinized blood from each donor separately and cultured at 37 °C with shaking at 200 rpm. To check the *S. Typhi* adaptation in human blood, samples were collected at five different time points (each time point con-

sisted of triplicate culture): time 0 (immediately after mixing bacteria with blood), and after 12, 24, 48 and 72 hours incubation at 37°C with shaking at 200 rpm and used for analysis by serial dilution as in case of *in vitro* growth analysis. Human blood from each donor without bacterial inoculum was used as negative control during viable bacterial cell enumeration experiment.

#### **Statistical analysis**

Two-way ANOVA with Bonferroni post-test was used for the statistical analysis. The statistical analysis was performed using GraphPadPrism5 with the significance set at a P of <0.05.

## **RESULTS**

Growth analysis of Vi-negative and Vi-positive *S. Typhi* isolates in TSB medium and human blood from four different donors over time course of 72 hours revealed that Vi-positive isolates have significantly shorter lag and generation times as compared to Vi-negative strains (Table 1). Vi-positive isolates grew at much faster rate as compared to Vi-negative isolates. In human whole blood, the lag time of Vi-negative isolates was doubled as compared to Vi-positive isolates. The viable cell density (CFU/ml) of *S. Typhi* capsulated strains (Vi-positive) was increased approximately 14 fold during 12 hours of incubation in blood. Contrasting growth pattern was observed in case of acapsulated isolates (Vi-negative). There was a fivefold (81 %) reduction in numbers after the first 12 hours of incubation in human blood (Figure 1B). They also showed different colonial morphology on MacConkey agar. Vi-positive isolates grew in the form of smooth circular colonies whereas Vi-negative isolates had circular colonies with rough boundaries having similar sizes (approximately 2-3 mm within 48 hours of incubation). On the whole, bacterial numbers increased approximately  $10^4$  (Vi-negative isolates) and  $10^5$  (Vi-positive isolates) over an incubation period of 72 hours in human blood. The bacterial count pattern for Vi-

positive isolates was similar in blood of four donors. It was also similar in case of Vi-negative isolates (Figure 2) indicating reproducibility of the results.

**Table 1:** Behavior of Vi-negative and Vi-positive isolates of *Salmonella* Typhi populations incubated at 37 °C in TSB and human blood

Growth curve values	Vi-	Vi+
Lag time (in hours) in TSB	10	8
Lag time (in hours) in blood	48	24
Generation time (in min) in TSB	34.73	12.72
Generation time (in min) in blood	102	72.24
Maximum population density (log-CFU/ml) in TSB	9.01±0.586	9.5±0.119
Maximum population density (log-CFU/ml) in blood	7.4±0.066	8.95±0.016
Colony morphology on MacConkey agar	Rough circular	Smooth circular

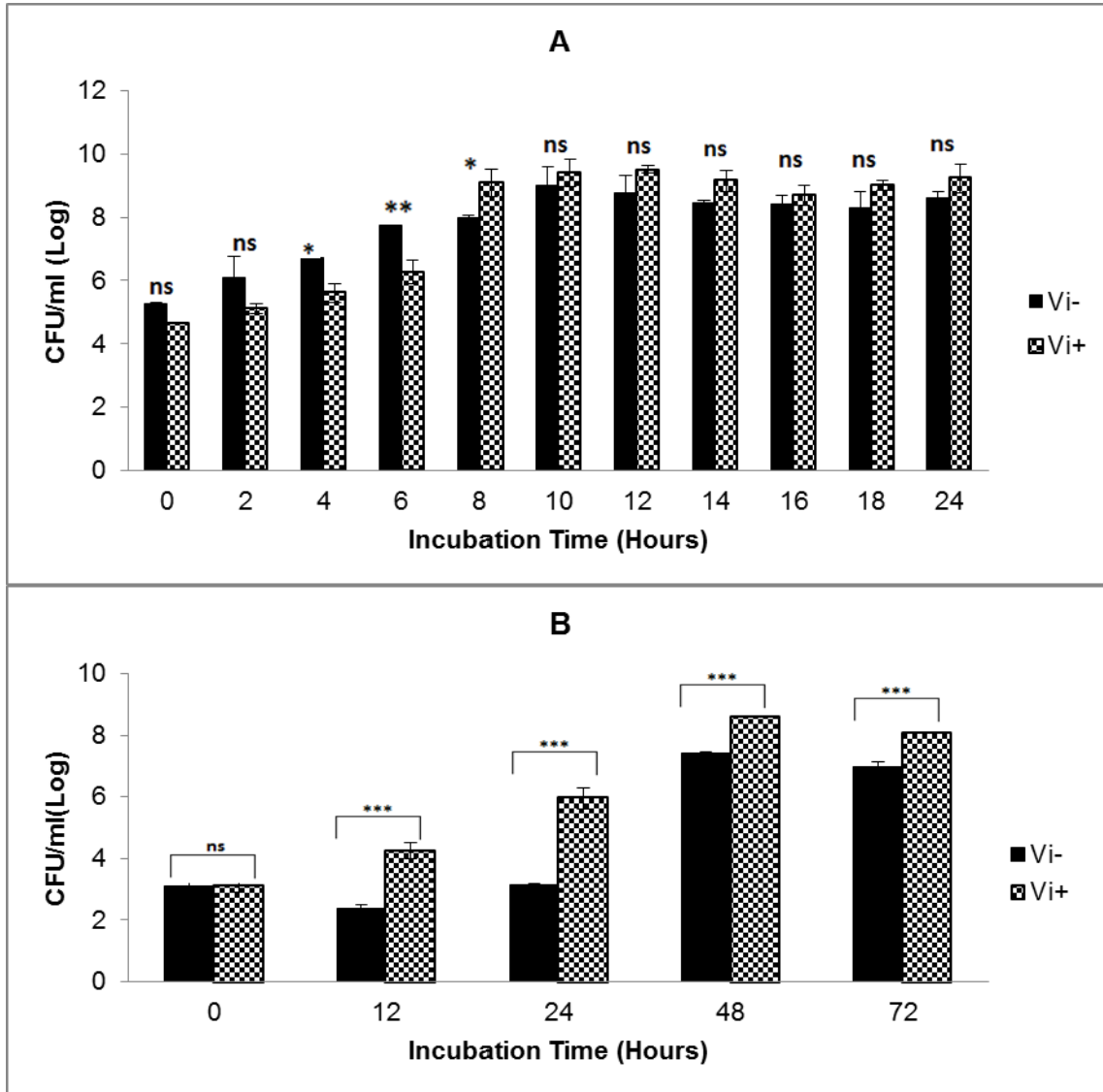
## DISCUSSION

Vi capsular polysaccharide has been known as a virulence factor of *S. Typhi* for the last several decades. This virulence antigen is expressed by *S. Typhi*, *Citrobacter freundii*, and some other serogroups of *Salmonellae*. In recent years, the old dogma that virulence of *S. Typhi* is essentially related to production of Vi antigen is shaken with the reports of Vi negative but pathogenic strains emerging from all over the world (Baker et al., 2005; Hone et al., 1988; Mehta and Arya, 2002; Pulickal et al., 2013; Wain et al., 2005). The potential of Vi negative strains to cause typhoid fever is now an established fact but the differences in the growth and survival of Vi negative and Vi positive variants during infection is still not clear.

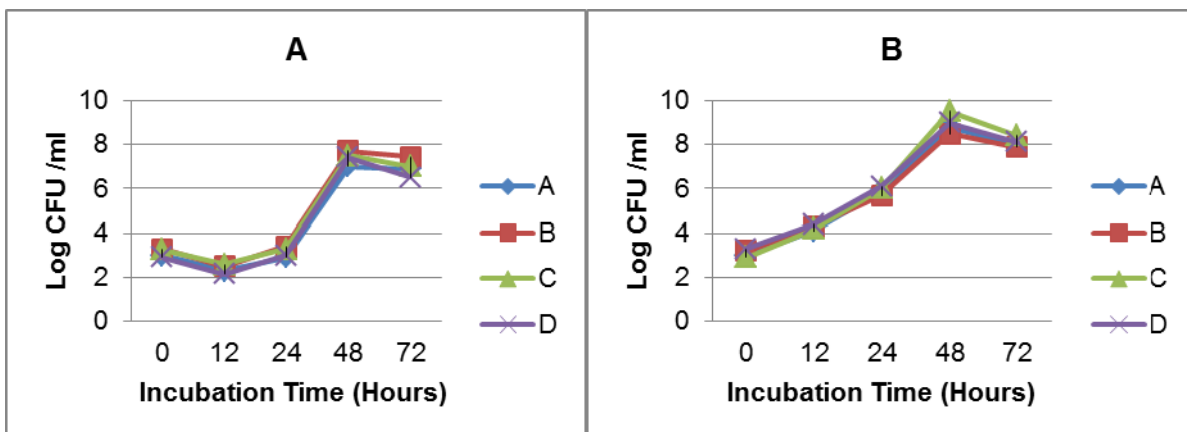
The Vi capsule is a protective component of *S. Typhi* and its role in intracellular survival of *S. Typhi* and in serum resistance has been recently reported (Hirose et al., 1997; Looney and Steigbigel, 1986). It is desirable to analyze the effect of absence of this protective antigen during natural infection. The comparative analysis of the growth patterns

of Vi-positive and Vi-negative *S. Typhi* isolates in a model mimicking conditions of natural infection can be a method of choice. In this study, human whole blood is selected as an *ex vivo* model because of its ability to mimic the behavior of *S. Typhi* growth in host, in response to cellular and humoral bactericidal mechanisms.

Considering previous reports about serum resistance in the presence of capsular Vi antigen (Bravo et al., 2008; Looney and Steigbigel, 1986), we investigated effect of the presence of Vi antigen on the bacterial survival and growth in human whole blood. Our results showed that presence of capsule affects growth of bacteria in human blood because capsulated isolates grow at a faster rate as compared to acapsulated isolates and they maintain sustainable growth. The lag time of Vi-negative isolates was double as compared to Vi-positive isolates. In case of Vi negative isolates, live cell density (CFU/ml) decreased about five fold within the first 12 hours of incubation in blood. It means that acapsulated isolates take longer time to adapt to blood environment where as capsulated isolates promptly acclimatize themselves due to presence of additional protective tool in the form of capsule. This is the major difference in survival and growth of Vi-negative and Vi-positive isolates in human blood as compared to TSB medium (Figure 1A, B). Therefore, it may be concluded that absence of capsule affects the survival and growth of Vi-negative isolates in human blood. Additionally, as expected from the analysis of blood from four different volunteers, overall survival pattern of both types of *S. Typhi* isolates remained consistent (Figure 2). Thus our results do not agree with a previous study indicating that presence of Vi capsule does not affect bacterial resistance to factors present in human blood (Bravo et al., 2008). Our results showed a marked difference in survival and growth of capsulated and acapsulated isolates in blood within the first 12 hours of incubation.



**Figure 1:** Growth of Vi-negative and Vi-positive *S. Typhi* isolates in TSB medium (A) and human whole blood(B)



**Figure 2:** Growth of Vi-negative (A) and Vi-positive (B) *S. Typhi* isolates in human whole blood from four donors

Looney and Steigbigel (1986) found that Vi-negative *S. typhi* isolates were extremely sensitive to non immune human serum and more than 99 % of bacterial cells were lysed within 40 minutes of their exposure to human serum. Our results support this report, although we found only 81 % reduction in viable Vi-negative bacteria during the first 12 hours of incubation in human blood. Another report by Hirose et al. (1997) pointed out that both types of *S. Typhi* strains can survive in resting macrophages although growth of acapsulated strains was inhibited. Neither Vi-positive nor Vi-negative *S. Typhi* survived in the activated human macrophages and only Vi-positive *S. Typhi* replicated inside resting macrophages as compared to Vi-negative strain (Hirose et al., 1997). Our results support this finding and highlight the protective role of Vi capsule.

In brief, we can say that absence of capsule does not only affect survival but also retard bacterial growth especially during the first 12 hours of incubation in human blood. It is probable that Vi-negative bacteria use some alternative strategies to protect themselves. To best of our knowledge, this is the first report regarding comparative growth analysis of Vi-positive and Vi-negative *S. Typhi* in whole human blood. Thus it can be concluded from the study that the absence of Vi capsular polysaccharide affects the survival of bacteria within human whole blood transiently but can critically affect its growth rate during the course of infection.

## REFERENCES

Baker S, Sarwar Y, Aziz H, Haque A, Ali A, Dougan G, et al. Detection of Vi-negative *Salmonella enterica* serovar typhi in the peripheral blood of patients with typhoid fever in the Faisalabad region of Pakistan. *J Clin Microbiol.* 2005;43:4418-25.

Bravo D, Silva C, Carter JA, Hoare A, Alvarez SA, Blondel CJ, et al. Growth-phase regulation of lipopolysaccharide O-antigen chain length influences serum resistance in serovars of *Salmonella*. *J Med Microbiol.* 2008;57:938-46.

Bueno SM, Santiviago CA, Murillo AA, Fuentes JA, Trombert AN, Rodas PI, et al. Precise excision of the large pathogenicity island, SPI7, in *Salmonella enterica* serovar Typhi. *J Bacteriol.* 2004;186:3202-13.

Crawford RW, Wangdi T, Spees AM, Xavier MN, Tsolis RM, Baumler AJ. Loss of very-long O-antigen chains optimizes capsule-mediated immune evasion by *Salmonella enterica* serovar Typhi. *MBio.* 2013;4(4):e00232-13.

Echenique-Rivera H, Muzzi A, Del Tordello E, Seib KL, Francois P, Rappuoli R, et al. Transcriptome analysis of *Neisseria meningitidis* in human whole blood and mutagenesis studies identify virulence factors involved in blood survival. *PLoS Pathog.* 2011;7:e1002027.

Graham MR, Virtaneva K, Porcella SF, Barry WT, Gowen BB, Johnson CR, et al. Group A *Streptococcus* transcriptome dynamics during growth in human blood reveals bacterial adaptive and survival strategies. *Am J Pathol.* 2005;166:455-65.

Haneda T, Winter SE, Butler BP, Wilson RP, Tukel C, Winter MG, et al. The capsule-encoding *viaB* locus reduces intestinal inflammation by a *Salmonella* pathogenicity island 1-independent mechanism. *Infect Immun.* 2009;77:2932-42.

Hirose K, Ezaki T, Miyake M, Li T, Khan AQ, Kawamura Y, et al. Survival of Vi-capsulated and Vi-deleted *Salmonella typhi* strains in cultured macrophage expressing different levels of CD14 antigen. *FEMS Microbiol Lett.* 1997;147:259-65.

Hone DM, Attridge SR, Forrest B, Morona R, Daniels D, LaBrooy JT, et al. A *galE* via (Vi antigen-negative) mutant of *Salmonella typhi* Ty2 retains virulence in humans. *Infect Immun.* 1988;56:1326-33.

Liu GR, Liu WQ, Johnston RN, Sanderson KE, Li SX, Liu SL. Genome plasticity and ori-ter rebalancing in *Salmonella typhi*. *Mol Biol Evol.* 2006;23:365-71.

Looney RJ, Steigbigel RT. Role of the Vi antigen of *Salmonella typhi* in resistance to host defense in vitro. *J Lab Clin Med.* 1986;108:506-16.

Massi MN, Shirakawa T, Gotoh A, Bishnu A, Hatta M, Kawabata M. Quantitative detection of *Salmonella enterica* serovar Typhi from blood of suspected typhoid fever patients by real-time PCR. *Int J Med Microbiol.* 2005;295:117-20.

Mehta G, Arya SC. Capsular Vi polysaccharide antigen in *Salmonella enterica* serovar typhi isolates. *J Clin Microbiol.* 2002;40:1127-8.

- Mereghetti L, Sitkiewicz I, Green NM, Musser JM. Extensive adaptive changes occur in the transcriptome of *Streptococcus agalactiae* (group B streptococcus) in response to incubation with human blood. *PLoS One*. 2008;3:e3143.
- Nair S, Alokam S, Kothapalli S, Porwollik S, Proctor E, Choy C, et al. *Salmonella enterica* serovar Typhi strains from which SPI7, a 134-kilobase island with genes for Vi exopolysaccharide and other functions, has been deleted. *J Bacteriol*. 2004;186:3214-23.
- Pulickal AS, Callaghan MJ, Kelly DF, Maskey M, Mahat S, Hamaluba M, et al. Prevalence and genetic analysis of phenotypically Vi-negative *Salmonella typhi* isolates in children from Kathmandu, Nepal. *J Trop Pediatr*. 2013;59:317-20.
- Raffatellu M, Santos RL, Chessa D, Wilson RP, Winter SE, Rossetti CA, et al. The capsule encoding the *viaB* locus reduces interleukin-17 expression and mucosal innate responses in the bovine intestinal mucosa during infection with *Salmonella enterica* serotype Typhi. *Infect Immun*. 2007;75:4342-50.
- Scharfman A, Kroczyński H, Carnoy C, Van Brussel E, Lamblin G, Ramphal R, et al. Adhesion of *Pseudomonas aeruginosa* to respiratory mucins and expression of mucin-binding proteins are increased by limiting iron during growth. *Infect Immun*. 1996;64:5417-20.
- Sizemore DR, Elsinghorst EA, Eck LC, Branstrom AA, Hoover DL, Warren RL, et al. Interaction of *Salmonella typhi* strains with cultured human monocyte-derived macrophages. *Infect Immun*. 1997;65:309-12.
- Sleator RD, Clifford T, Hill C. Gut osmolarity: a key environmental cue initiating the gastrointestinal phase of *Listeria monocytogenes* infection? *Med Hypotheses*. 2007;69:1090-2.
- Tartera C, Metcalf ES. Osmolarity and growth phase overlap in regulation of *Salmonella typhi* adherence to and invasion of human intestinal cells. *Infect Immun*. 1993;61:3084-9.
- Toledo-Arana A, Dussurget O, Nikitas G, Sesto N, Guet-Revillet H, Balestrino D, et al. The *Listeria* transcriptional landscape from saprophytism to virulence. *Nature*. 2009;459:950-6.
- Tran QT, Gomez G, Khare S, Lawhon SD, Raffatellu M, Baumlér AJ, et al. The *Salmonella enterica* serotype Typhi Vi capsular antigen is expressed after the bacterium enters the ileal mucosa. *Infect Immun*. 2010;78:527-35.
- Wain J, Pham VB, Ha V, Nguyen NM, To SD, Walsh AL, et al. Quantitation of bacteria in bone marrow from patients with typhoid fever: relationship between counts and clinical features. *J Clin Microbiol*. 2001;39:1571-6.
- Wain J, House D, Zafar A, Baker S, Nair S, Kidgell C, et al. Vi antigen expression in *Salmonella enterica* serovar Typhi clinical isolates from Pakistan. *J Clin Microbiol*. 2005;43:1158-65.
- Wangdi T, Lee CY, Spees AM, Yu C, Kingsbury DD, Winter SE, et al. The Vi capsular polysaccharide enables *Salmonella enterica* serovar typhi to evade microbe-guided neutrophil chemotaxis. *PLoS Pathog*. 2014;10:e1004306.
- Wilson RP, Winter SE, Spees AM, Winter MG, Nishimori JH, Sanchez JF, et al. The Vi capsular polysaccharide prevents complement receptor 3-mediated clearance of *Salmonella enterica* serotype Typhi. *Infect Immun*. 2011;79:830-7.
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